

---

Biological Report 90(5)  
June 1990

## **Ecology and Assessment of Warmwater Streams: Workshop Synopsis**



---

Fish and Wildlife Service  
**U.S. Department of the Interior**

DISTRIBUTION STATEMENT A

Approved for public release;  
Distribution Unlimited

PRINTED QUALITY DOCUMENTATION

## ***Biological Report***

This publication series of the Fish and Wildlife Service comprises reports on the results of research, developments in technology, and ecological surveys and inventories of effects of land-use changes on fishery and wildlife resources. They may include proceedings of workshops, technical conferences, or symposia; and interpretive bibliographies.

Copies of this publication may be obtained from the Publications Unit, U.S. Fish and Wildlife Service, 1849 C Street, N.W., Mail Stop 130—ARLSQ, Washington, DC 20240, or may be purchased from the National Technical Information Service (NTIS), 5285 Port Royal Road, Springfield, VA 22161.

**ISSN 0895-1926**

**This publication may be cited as follows:**

Bain, M. B., editor. 1990. Ecology and assessment of warmwater streams: workshop synopsis. U.S. Fish Wildl. Serv., *Biol. Rep.* 90(5). 44 pp.

**Chapters may be cited as follows:**

Neves, R. J. 1990. Biology and ecology of mollusks in streams. Pages 8–9 in M. B. Bain, ed. Ecology and assessment of warmwater streams: workshop synopsis. U.S. Fish Wildl. Serv., *Biol. Rep.* 90(5). 44 pp.

Biological Report 90(5)  
June 1990

# Ecology and Assessment of Warmwater Streams: Workshop Synopsis

Mark B. Bain

*U.S. Fish and Wildlife Service  
Alabama Cooperative Fish and Wildlife Research Unit  
Department of Fisheries and Allied Aquacultures  
Auburn University  
Auburn, Alabama 36849*

U.S. Department of the Interior  
Fish and Wildlife Service  
Washington, DC 20240

## Contents

|  | Page   |
|--|--------|
| Preface .....  | iv     |
| Acknowledgments .....  | v      |
| <br><b>Overview.....</b>   | <br>1  |
| Ecological Research and Impact Assessment: Complementary but Different Endeavors.<br><i>Mark B. Bain</i> .....   | 2      |
| <br><b>Ecological Research.....</b>  | <br>5  |
| Ecosystem Characteristics and Biological Productivity of Southeastern Coastal Plain<br>Blackwater Rivers. <i>Arthur C. Benke</i> .....                           | 6      |
| Biology and Ecology of Mollusks in Streams. <i>Richard J. Neves</i> .....  | 8      |
| Aquatic and Terrestrial Linkages: The Role of Floodplains. <i>J. Vaun McArthur</i> .....   | 10     |
| Ecology of Southeastern Stream Fishes: Geographic, Macro- and Microhabitat<br>Considerations. <i>Stephen T. Ross</i> .....                                       | 11     |
| A Pluralistic View of Factors Influencing Warmwater Stream Fishes: Implications for<br>Environmental Management. <i>Isaac J. Schlosser</i> .....                 | 14     |
| Fish Community Structure and Stability in Warmwater Midwestern Streams.<br><i>William J. Matthews</i> .....  | 16     |
| Centrarchid-Habitat Associations in Ozark Streams. <i>Charles F. Rabeni</i> .....  | 18     |
| Fish Larvae—Ecologically Distinct Organisms. <i>Darrel E. Snyder</i> .....   | 20     |
| <br><b>Assessment and Research.....</b>  | <br>25 |
| The IBI: A Quantitative, Easily Communicated Assessment of the Health and Complexity<br>of Entire Fish Communities. <i>Robert M. Hughes</i> .....                | 26     |
| Comparison of Instream Flow Methods for Western Virginia.<br><i>Donald J. Orth and Paul M. Leonard</i> .....   | 29     |
| Minimum Flow is a Myth. <i>Clair B. Stalnaker</i> .....  | 31     |
| Considerations in Applying IFIM to Warmwater Streams. <i>John M. Nestler</i> .....   | 34     |
| Determination of Instream Flow Needs at Hydroelectric Projects in the Northeast.<br><i>Gordon W. Russell</i> .....   | 36     |
| Discharge versus Habitat in Steep-gradient Piedmont Streams. <i>Jeffrey W. Foltz</i> .....   | 38     |
| Use of Multiple Methods for Instream Flow Recommendations—A State Agency<br>Approach. <i>Steven E. Reed and James S. Mead</i> .....                              | 40     |
| Stream Habitat Analysis and Instream Flow Assessment: A State–Federal Effort in<br>Arkansas. <i>Danny J. Ebert, Steve P. Filipek, and Kelly M. Russell</i> ..... | 43     |

## Preface

Natural resource agencies are frequently confronted by applications for permits and licenses for developments and actions that affect the aquatic resources of warmwater streams and rivers. At this time, there are no generally accepted methods for assessing impacts and developing mitigation measures for warmwater streams. To begin correcting these deficiencies, U.S. Fish and Wildlife Service biologists and their State agency colleagues need state-of-the-art knowledge on (1) warmwater stream ecology, emphasizing fish and habitat; and (2) existing methods for assessing stream quality and impact.

The need for current information on the ecology and assessment of warmwater streams emerged from the U.S. Fish and Wildlife Service field offices in the Southeast and their State agency colleagues. In response to this need, Warren T. Olds (Assistant Regional Director for Fish and Wildlife Enhancement) and William K. Seitz (Office of Information Transfer) decided in late 1987 that a workshop would be held for biologists involved in managing stream resources and assessing impacts of developments on streams and rivers. The scope and format of the workshop were determined in January 1988 by a steering committee composed of the following persons: Mark B. Bain, Alabama Cooperative Fish and Wildlife Research Unit; John W. Baumeister and James D. Brown, U.S. Fish and Wildlife Service Region 4 Office; Reis Collier, U.S. Fish and Wildlife Service Field Office—Raleigh, North Carolina; Larry E. Goldman, U.S. Fish and Wildlife Service Field Office—Daphne, Alabama; Sam Hamilton, U.S. Fish and Wildlife Service Office of Federal Activities; James Kirkwood, U.S. Fish and Wildlife Service Region 4 Office; and James Layzer, Tennessee Cooperative Fishery Research Unit.

The workshop was held at Lakepoint State Park Resort, Eufaula, Alabama, on 26–27 October 1988. The agenda of the workshop was composed of presentations by two groups of biologists: stream ecologists with extensive basic research experience, and applied research biologists experienced in assessing effects on warmwater streams. There was no intent to merge the basic and applied research; rather the aim was to contrast ecological knowledge with existing impact assessment methods. Biologists and ecologists were invited from Service offices in the Southeast, State natural resource agencies in 11 southeastern States, and Service research and regional offices nationwide. Workshop attendees were about equally divided among these three groups.

This synopsis, which follows the format of the workshop, provides an overview of the topics covered with a list of detailed information sources and expert contacts. Speakers summarized their main points and assembled a list of key papers and books that complement the material they covered or that contain information used in their presentation. Together, the summaries and information sources provide a beginning point for biologists who need current and detailed information on warmwater stream ecology and assessment methodologies.

## **Acknowledgments**

The Office of Information Transfer provided funding for the workshop and this synopsis. D. Parrotte and his staff at the Auburn University Continuing Education Office handled workshop arrangements and administration. The Aquatic Systems Branch of the National Ecology Research Center typed and assembled the first draft of this manuscript. Many subsequent versions were corrected and revised by J. Christian of the Alabama Cooperative Fish and Wildlife Research Unit. P. Angermeier, H. Bart, J. Boltz, C. Courter, and R. Krska reviewed the manuscript.

The workshop sponsors and I thank all speakers for contributing their time and effort to prepare oral presentations and written summaries. I especially appreciate the Fish and Wildlife Service speakers, all of whom chose to contribute to this effort using funding and time from their own programs. This cooperative attitude reflects an unusual degree of commitment by Service people.

# Overview

## Ecological Research and Impact Assessment: Complementary but Different Endeavors

by

Mark B. Bain

*U.S. Fish and Wildlife Service  
Alabama Cooperative Fish and Wildlife Research Unit  
331 Funchess Hall, Auburn University  
Auburn, Alabama 36849*

While both science and impact assessment rely on technical studies, they differ in fundamental goals, approach and scope, and final products. Research and management biologists come from similar educational backgrounds, share similar career interests, and have a high regard for biological knowledge. However, varied philosophy and interests emerge due to different work settings and responsibilities. Science is defined as systematized knowledge derived from observation, study, and experimentation carried on in order to determine the nature or principles of what is being studied. The basic approach to study is the scientific method: observation, hypothesis formation, hypothesis testing, deriving results, and interpreting findings relative to principle or theories. Impact assessment is the process of documenting the important consequences of proposed actions by (1) objective analyses of current and predicted conditions and (2) subjective evaluation of the significance of predicted changes. In contrast to the scientific method, the impact assessment method uses distinctly different steps: reviewing proposed actions, documenting baseline conditions, identifying possible impacts, predicting changes, documenting significant impacts, and formulating recommendations. The definitions and methods of both science and impact assessment reflect the fundamental differences between these endeavors.

Other aspects of science and assessment also distinguish the professions. Most notable among these is the interpretation of the term *significance*. To scientists, the term refers to the probability level associated with rejecting a true null hypothesis; in practice, a result is significant if the data deviate enough (probability of occurrence  $\leq 0.05$ ) from the prediction of the null hypothesis to be considered different. To assessment biologists, significance is determined by a subjective determination that in

practice depends largely on agency policies, public concerns, legal standards, personal preferences, and past case histories. The scope of study differs between the professions. Scientists work hard to narrow the number of factors affecting their observations, whereas assessment studies attempt to assemble a diverse array of information, data, and past agency actions to defend a recommendation. Science progresses through the peer-reviewed literature, advancement and replacement of theories and principles, and university education programs. Impact assessment lacks modes of progress due to the limited nature of professional impact assessment journals, dependence on changing agency policies and laws, lack of formal educational programs, and low emphasis on disseminating assessment study findings.

Although science and assessment differ in many ways, the professions complement one another and benefit from exchange of needs and information. Natural resource agencies need reliable and accepted information, methods, and principles from scientists. Most important are the key factors, processes, mechanisms, and structural properties that represent the essential characteristics and functions of species and biological systems. In addition, when this type of information is reduced to accepted basics, science is directly serving natural resource agency biologists. To enhance the value of science to natural resource agencies, assessment biologists should convey information on the type of scientific studies that are most needed: intensive studies of a few factors or extensive studies of complex system patterns, key biological levels of study (species to ecosystems), geographic scale, habitat types, and duration (short- or long-term). Each profession benefits by understanding the difference and needs of the other so that information can be exchanged effectively for mutual benefit.

## Information Sources

- Barnthouse, L. W., J. Boreman, S. W. Christensen, C. P. Goodear, W. Van Winkle, and D. S. Vaughan. 1984. Population biology in the courtroom: the Hudson River controversy. *BioScience* 34:14-19.
- Beanlands, G. E., and P. N. Duinker. 1984. An ecological framework for environmental impact assessment. *J. Environ. Manage.* 18:267-277.
- Cairns, J., Jr. 1988. Politics, economics, science—going beyond disciplinary boundaries to protect aquatic ecosystems. Pages 1-16 in M. S. Evans, ed. *Toxic contaminants and ecosystem health: a Great Lakes focus*. John Wiley & Sons, Inc., New York.
- Haug, P. T., R. W. Burwell, A. Stein, and B. L. Bandurski. 1984. Determining the significance of environmental issues under the National Environmental Policy Act. *J. Environ. Manage.* 18:15-24.
- Hecky, R. E., R. W. Newbury, R. A. Bodaly, K. Patalas, and D. M. Rosenburg. 1984. Environmental impact prediction and assessment: the Southern Indian Lake experience. *Can. J. Fish. Aquat. Sci.* 41:720-732.
- Larkin, P. A. 1984. A commentary on environmental impact assessment for large projects affecting lakes and streams. *Can. J. Fish. Aquat. Sci.* 41:1121-1127.
- Norton, G. A., and B. H. Walker. 1982. Applied technology: towards a positive approach I. The context of applied ecology. *J. Environ. Manage.* 14:309-324.
- Orians, G. H. 1986. Ecological knowledge and environmental problem-solving. Committee on the Application of Ecological Theory to Environmental Problems, U.S. National Research Council, National Academy Press, Washington, D.C.
- Rosenburg, D. M., and 14 coauthors. 1981. Recent trends in environmental impact assessment. *Can. J. Fish. Aquat. Sci.* 38:59-624.
- Toth, E. F., and J. W. Baglien. 1986. When habitats fail as predictors—the manager's viewpoint. Pages 255-256 in J. Verner, M. L. Morrison, and C. J. Ralph, eds. *Wildlife 2000*. The University of Wisconsin Press, Madison. 454 pp.
- Van Horn, B. 1986. When habitats fail as predictors—the researcher's viewpoint. Pages 257-260 in J. Verner, M. L. Morrison, and C. J. Ralph, eds. *Wildlife 2000*. The University of Wisconsin Press, Madison. 454 pp.
- Walker, B. H., and G. A. Norton. 1982. Applied ecology: towards a positive approach II. Applied ecological analysis. *J. Environ. Manage.* 14:325-342.

# Ecological Research

Presentations by stream ecologists address a variety of organisms, physical characteristics, and temporal properties of stream systems. Despite the wide differences in research topics and background of speakers, four recurrent themes are evident. First, the need to understand the extent and causes of natural variation in biological communities is stressed as a prerequisite for developing effective management methods. Many studies have identified that populations, trophic organization, interspecific interactions, and species compositions are dynamic and change through time in stream systems. The magnitude of natural change is recognized as considerable and difficult to predict but not solely a random phenomenon. Second, the importance of natural variability and timing of flow events (especially floods and floodplain inundation) is recognized. Much of the variability in stream biota probably comes from the natural variability in streamflows. Research shows that organisms are adapted to seasonal cycles in stream discharge, and maintaining these cycles will be necessary to preserve natural system properties. Third, the critical importance of early life stages in determining adult population dynamics is discussed. Although several speakers identified this area as integral for explaining the natural dynamics of stream fish populations, they also agree that present knowledge of early life biology is poor and hampered by the continued emphasis on studies of adults and late juveniles. Finally, the complexity observed in warmwater communities and species-habitat relations is emphasized. Despite high complexity, the presentations include some generalized patterns (e.g., invertebrate production, faunal declines, fish-habitat organizations) that are useful for stream management. In general, recent research and probable future emphasis seem to center on relations among different types of aquatic habitats, causes of variance in the biological characteristics of streams, and the role of temporal habitat variability in communities.

## Ecosystem Characteristics and Biological Productivity of Southeastern Coastal Plain Blackwater Rivers

by

Arthur C. Benke

*Department of Biology  
University of Alabama  
Tuscaloosa, Alabama 35487*

Blackwater rivers in the Coastal Plain of the southeastern United States are characterized by broad floodplain swamp forests and the presence of large amounts of woody debris (snags) in the main channel. Since they are located in a relatively flat landscape with low gradient, Coastal Plain streams are not as subject to impoundment as streams and rivers in higher gradients. However, snagging, channelization, and drainage of adjacent wetlands occur frequently in such systems, and all three can cause substantial ecological damage.

The Satilla and Ogeechee rivers are blackwater river systems in the southeastern Coastal Plain. They are the two largest river systems in Georgia that remain free from any major impoundments from their headwaters to the sea. Furthermore, there have not been major attempts to clear snags or channelize most of the upper portions of these rivers. However, the lower section of the Satilla was regularly cleared of snags until the early 1950's.

In both the Satilla and Ogeechee rivers, snags are the major site of invertebrate biomass, production, and diversity (Benke et al. 1984a, 1984b; Wallace and Benke 1984; Benke and Meyer 1988). River sediments generally have lower production and diversity, although they constitute larger areas. Many of the snag animals are filter-feeding aquatic insects (e.g., caddisflies, black flies, midges, mayflies). Other snag-dwellers include invertebrate predators such as dragonflies, stoneflies, and hellgrammites. Furthermore, the majority of drifting invertebrates (70–80% of numbers and biomass) originates from the snag habitat in the Satilla River (Benke et al. 1986). Also in the Satilla River, at least 50% of the prey of insectivorous fishes (e.g., bluegill, redbreast) originates from snags. Thus, invertebrate production—and hence probably much fish production—is directly related to the abundance of snags.

Interactions with the adjacent river swamps (increasing river width by as much as 40 times) are

extremely important to animal productivity in these Coastal Plain rivers. The major organic foods for snag invertebrates seem to originate from swamps in the form of suspended soil bacteria or fine particulate organic matter (Edwards 1987; Edwards and Meyer 1987; Wallace et al. 1987). During flood periods, invertebrate production increases on the submerged swamp substrate and on wetted tree trunks. Inundation of the floodplain usually occurs for 3–4 months of the year and provides for a great expansion of feeding and spawning opportunities for many fish species.

Long-term management of Coastal Plain rivers should emphasize protection of the floodplain forests, especially by discouraging snag removal and channelization. In areas that have been largely cleared of snags or channelized, fish and wildlife managers should consider the introduction of woody debris to reestablish productive invertebrate and fish habitat.

### Information Sources

- Benke, A. C., R. L. Henry, D. M. Gillespie, and R. J. Hunter. 1985. Importance of the snag habitat for animal production in a southeastern stream. *Fisheries* 10:8–13.
- Benke, A. C., R. J. Hunter, and F. K. Parrish. 1986. Invertebrate drift dynamics in a subtropical blackwater river. *J. North Am. Benthol. Soc.* 5:173–190.
- Benke, A. C., and J. L. Meyer. 1988. Structure and function of a blackwater river in southeastern USA. *Proc. Int. Assoc. Theor. Appl. Limnol.* 23:1209–1218.
- Benke, A. C., T. C. Van Arsdall, D. M. Gillespie, and F. K. Parrish. 1984a. Invertebrate productivity in a subtropical blackwater river: the importance of habitat and life history. *Ecol. Monogr.* 54:25–63.
- Benke, A. C., T. C. Van Arsdall, D. M. Gillespie, and F. K. Parrish. 1984b. Invertebrate productivity in a subtropical blackwater river. *BioScience* 34:443–444.
- Edwards, R. T. 1987. Sestonic bacteria as a food source for filtering invertebrates in two southeastern blackwater rivers. *Limnol. Oceanogr.* 32:221–234.

- Edwards, R. T., and J. L. Meyer. 1987. Bacteria as a food source for black fly larvae in a blackwater river. *J. North Am. Benthol. Soc.* 6:241-250.
- Harmon, M. E., J. F. Franklin, F. J. Swanson, P. Sollins, S. V. Gregory, J. D. Lattin, N. H. Anderson, S. P. Cline, N. G. Aumen, and J. R. Sedell. 1986. Ecology of coarse woody debris in temperate ecosystems. *Adv. Ecol. Res.* 15:133-302.
- Marzolf, G. R. 1978. The potential effects of clearing and snagging on stream ecosystems. U.S. Fish Wildl. Serv., FWS/OBS-78/14. 31 pp.
- Sedell, J. R., and J. L. Froggatt. 1984. Importance of streamside forests to large rivers: the isolation of the Willamette River, Oregon, USA, from its floodplain by snagging and streamside forest removal. *Proc. Int. Assoc. Theor. Appl. Limnol.* 22:1828-1834.
- Triska, F. J. 1984. Role of wood debris in modifying channel geomorphology and riparian areas of a large lowland river under pristine conditions: a historical case study. *Proc. Int. Assoc. Theor. Appl. Limnol.* 22:1876-1892.
- Wallace, J. B., and A. C. Benke. 1984. Quantification of wood habitat in subtropical Coastal Plain streams. *Can. J. Fish. Aquat. Sci.* 41:1643-1652.
- Wallace, J. B., A. C. Benke, A. H. Lingle, and K. Parson. 1987. Trophic pathways of macroinvertebrate primary consumers in subtropical blackwater streams. *Arch. Hydrobiol. Suppl.* 74:423-451.

## Biology and Ecology of Mollusks in Streams

by

Richard J. Neves

*U.S. Fish and Wildlife Service  
Virginia Cooperative Fish and Wildlife Research Unit  
Virginia Polytechnic Institute and State University  
Blacksburg, Virginia 24061*

The indigenous freshwater mollusk fauna of the United States consists of about 765 species, including 500 species of snails, 40 species of fingernail clams, and 225 species of mussels. Two major subgroups of snails, pulmonates (lunged) and prosobranchs (gilled), have adapted to most freshwater environments. Pulmonate snails lack an operculum, respire via a pulmonary sac, are hermaphroditic (monoecious), and form light shells for a fairly active existence. In contrast, prosobranch snails have an operculum, respire through gills, have separate sexes, and form heavier shells for a more sedentary lifestyle. The 15 families of snails in freshwater rivers in the southeastern United States are dominated by the Pleuroceridae. These snails are small to medium in size and have distinctive solid, dextral shells, which are used for species identification. Pleurocerids are most abundant on cobbles and boulders in rocky shoals.

Freshwater fingernail clams (Pisidiidae) and the exotic asiatic clam (*Corbicula fluminea*) are small bivalves (<35 mm) that occupy both lotic and lentic habitats. Pisidiids are hermaphroditic, reproduce once or twice each year, and brood their young in marsupial sacs. The asiatic clam is also hermaphroditic, spawning twice a year and brooding embryos in its gills. It lives 2–3 years and achieves a maximum size of 35 mm in most rivers. Since 1924, this species has spread from the Pacific Northwest and now occurs in roughly 40 States. Because of its high reproductive potential, fast growth rate, short generation time, and high population densities, this exotic clam may compete with—and eventually displace—some of the indigenous bivalve fauna.

Of the 225 freshwater mussel species (Unionidae) in the United States, 34 are listed by the Federal Government as endangered; most of these occur in the Southeast, and many more species have been proposed for listing. The mussel reproductive cycle

is similar for all species. During the spawning period, males release sperm into the water column. The sperm are then taken in by females during siphoning. Eggs are fertilized and incubated in the gills until larvae (glochidia) are mature. Glochidia are released into the water and must attach to the fins or gills of appropriate host fishes to encyst and metamorphose to the free-living juvenile stage. Juveniles are about 200 µm and require roughly 5 years to become sexually mature. Mussels are the longest-lived freshwater mollusks; individuals of some species have been aged at more than 50 years. The extreme longevity and the annual release of prodigious numbers (10,000–100,000) of glochidia per female provide the necessary reproductive potential to contact appropriate fish hosts and continue recruitment.

Habitat requirements for most unionids are fairly specific: clean, flowing water and a stable substratum of mixed particle sizes. In the Southeast, declines in the mussel fauna have been attributed principally to river impoundments, siltation, and water pollution. Numerous hydroelectric and flood control reservoirs have eliminated riverine conditions in most drainages, altering substrate and changing fish species composition. Poor land-use and mining practices have silted many streams and rivers, eliminating or reducing mussel populations in many river reaches. Point and nonpoint pollution have resulted in acute and chronic mortalities from herbicides, toxic spills, heavy metal discharges, sewage plant effluents, coal waste deposition, and many other anthropogenic activities. Over the last decade, an improvement in water quality in many rivers has occurred, which may allow the natural recolonization and recovery of some species. However, most endangered species are not likely to recover without the implementation of specific activities identified in recovery plans. Additional research and improvements in environmental quality

should provide opportunities to expedite the recovery of these unionids.

## Information Sources

- Basch, P. F. 1963. A review of the recent freshwater limpet snails of North America (Mollusca:Pulmonata). *Bull. Mus. Comp. Zool. Harv. College* 129(8):399-461.
- Beetle, D. E. 1973. A checklist of the land and freshwater mollusks of Virginia. *Sterkiana* 49:21-35.
- Branson, B. A. 1970. Checklist and distribution of Kentucky aquatic gastropods. *Ky. Fish. Bull.* 54:1-20.
- Burch, J. B. 1972. Freshwater sphaeriacean clams (Mollusca:Pelecypoda) of North America. U.S. Environ. Prot. Agency, Biota of Freshwater Ecosystems, Identification Manual 3. 31 pp.
- Burch, J. B. 1973. Freshwater unionacean clams (Mollusca:Pelecypoda) of North America. U.S. Environ. Prot. Agency, Biota of Freshwater Ecosystems, Identification Manual 11. 176 pp.
- Burch, J. B. 1982. Freshwater snails (Mollusca:Gastropoda) of North America. U.S. Environ. Prot. Agency EPA 600/3-82-026. 294 pp.
- Clench, W. J., and R. D. Turner. 1956. Freshwater mollusks of Alabama, Georgia, and Florida from the Escambia to the Suwanee River. *Bull. Fla. State Mus.* 1:97-239.
- Dawley, C. 1965. Checklist of freshwater mollusks of North Carolina. *Sterkiana* 19:35-39.
- Dundee, D. S. 1974. Catalog of introduced mollusks of eastern North America (north of Mexico). *Sterkiana* 55:1-37.
- Mazyck, W. G. 1913. Catalog of the mollusca of South Carolina. *Contrib. Charleston Mus.* 2:1-39.
- Neves, R. J. 1987. Recent die-offs of freshwater mussels in the United States: an overview. Pages 7-18 in R. J. Neves, ed. Proceedings of the workshop on die-offs of freshwater mussels in the United States. Virginia Tech Press, Blacksburg. 166 pp.
- Neves, R. J., and S. N. Moyer. 1988. Evaluation of techniques for age determination of freshwater mussels (Unionidae). *Am. Malacol. Bull.* 6:179-188.
- Neves, R. J., and J. C. Widlak. 1987. Habitat ecology of juvenile freshwater mussels (Bivalvia:Unionidae) in a headwater stream in Virginia. *Am. Malacol. Bull.* 5:1-7.
- Shoup, C. S. 1974. A bibliography of the zoology of Tennessee and the Tennessee Valley region. U.S. Atomic Energy Commission, Technical Information Center, Oak Ridge, Tenn. 251 pp.
- Thompson, F. G. 1968. The aquatic snails of the family Hydrobiidae of peninsular Florida. University of Florida Press, Gainesville. 268 pp.
- Thompson, F. G. 1984. The freshwater snails of Florida: a manual for identification. University of Florida Press, Gainesville. 94 pp.
- Zale, A. V., and R. J. Neves. 1982. Reproductive biology of four freshwater mussel species (Mollusca:Unionidae) in Virginia. *J. Freshwater Invertebr. Biol.* 1:17-28.
- Zale, A. V., and R. J. Neves. 1982. Fish hosts of four species of lampsiline mussels (Mollusca:Unionidae) in Big Moccasin Creek, Virginia. *Can. J. Zool.* 60:2535-2542.

## Aquatic and Terrestrial Linkages: The Role of Floodplains

by

J. Vaun McArthur

*Savannah River Ecology Laboratory*

*Drawer E*

*Aiken, South Carolina 29801*

Linkages between terrestrial and aquatic ecosystems occur at several orders of scale. At a landscape scale, stream basin characteristics control longitudinal and lateral (floodplain) interactions among aquatic habitats. The amount of organic matter input from the basin is dependent on the shape, recent use history, and nature of the floodplain. Lateral movement of organic matter has been severely affected in many river systems by levee districts, which have effectively blocked the stream from its valley. The nature and timing of organic matter input into lotic ecosystems will directly affect processing this material. Changes in the species composition of the riparian forest will affect the invertebrate assemblages in the stream because invertebrate life histories have evolved to respond to the resources that have historically been available. Changes in the invertebrate community can directly affect the fish assemblages. Removal of snag habitat has been a major cause in the decoupling of terrestrial and aquatic interactions. Debris dams serve as habitat for fish and provide sites for accumulation and subsequent processing of organic matter.

The closest link between terrestrial and aquatic ecosystems is bacteria. It has been shown that bacteria can only respond to natural sources of organic matter to which they have been previously exposed. This response is both physiologic and genetic. Populations of bacteria appear to be adapted to the organic matter found in the adjacent riparian and floodplain habitats. By changing the source of organic matter, the aquatic systems become less efficient in processing, and the greater portion of material is lost. As processing efficiency

declines, there is less secondary production of invertebrates, which will lead to a decrease in fish and other vertebrates that rely on the invertebrate biomass.

More research is needed to determine how tight the linkage between terrestrial and aquatic systems is and how changes affect higher trophic levels. Clearly, management of floodplain and riparian forests will affect all trophic levels in stream systems, and it is important that we document these effects so that effective management practices can be developed.

## Information Sources

- McArthur, J. V. 1989. Aquatic and terrestrial linkages: floodplain functions. In D. D. Hook and R. Lea, eds. Proceedings of the symposium: the forested wetlands of the southern United States. U.S. For. Serv. Gen. Tech. Rep. SE-50.
- McArthur, J. V., D. A. Kovacic, and M. H. Smith. 1988. Genetic diversity in natural populations of a soil bacterium across a landscape gradient. Proc. Nat. Acad. Sci. 85:9621-9624.
- Minshall, G. W. 1988. Stream ecosystem theory: a global perspective. J. North Am. Benthol. Soc. 7:263-288.
- Minshall, G. W., R. C. Petersen, K. W. Cummins, T. L. Bott, J. R. Sedell, C. E. Cushing, and R. L. Vannote. 1983. Interbiome comparison of stream ecosystem dynamics. Ecol. Monogr. 53:1-25.
- Naiman, R. J., H. Decamps, J. Pastor, and C. A. Johnston. 1988. The potential importance of boundaries to fluvial ecosystems. J. North Am. Benthol. Soc. 7:289-306.
- Webster, J. R., and E. F. Benfield. 1986. Vascular plant breakdown in freshwater ecosystems. Annu. Rev. Ecol. Syst. 17:567-594.

## Ecology of Southeastern Stream Fishes: Geographic Macro- and Micro-habitat Considerations

by

Stephen T. Ross

*Department of Biological Sciences  
University of Southern Mississippi  
Hattiesburg, Mississippi 39406*

An understanding of natural variation in fish assemblage composition is important in developing and implementing stream management plans. Over the last decade, fish ecologists have worked to understand the patterns of organization and variation in communities and then to explore the mechanisms responsible for these patterns. Studies of stream fish assemblages necessarily emphasize current conditions and interactions, simply because we are faced with finite lifespans, careers, and budgets. While human and professional mortality are facts that we must deal with, our mortality—and the resulting short-term view of ecological communities—has led to problems of interpretation. I will identify three problems confronting fish community ecologists and suggest approaches to dealing with them.

The first problem is that ecologists often approach communities with the idea that biotic processes that structured such systems are still ongoing and are unaltered in the particular habitat under study—the fallacy of community stasis. It is important to have a temporal perspective on the subjects that we study as fish ecologists—namely species, assemblages, river systems, populations, and microhabitats. In terms of species, the age of most of the fish fauna in the southeastern United States probably dates from the late Miocene or early Pliocene. For instance, ancestral suckers, minnows, and darters are thought to have dispersed over a North American land route (Beringia) during the early Tertiary (Gilbert 1976), when such a connection between Europe and America still existed.

Somewhat later, the modern darter (tribe *Etheostomatini*) arose from an unknown percid ancestor. Vicariant events, such as isolation in many small streams and the disruption of highland areas, led to the evolution of numerous species of *Etheostoma* and *Percina* (Collette and Banarescu 1977). The minnows date from about the same age, with diversification of the largest genus, *Notropis*, probably

occurring in the Mississippi Basin in the early to middle Pliocene (Gilbert 1964, for *Luxilus*). Fossil data indicate that centrarchids may date from the Oligocene (Gilbert 1976), and *Lepomis* occurred in the Miocene (Swift and Wing 1968). Thus, the three largest families of freshwater fishes have diverged since the Miocene or Pliocene (Gilbert 1964; Collette and Banarescu 1977), and therefore we are most concerned with recent, Pleistocene, and Pliocene geology in attempting to understand fish distributions and the history of interactions.

In dealing with the diverse fishes of the North American highlands, which include the area of greatest diversity of North American freshwater fishes, Mayden (1987a, 1987b) concluded that these faunas are older—probably much older—than the Pleistocene. While actual dates of origin of individual species are uncertain and will probably remain so, it is evident that the organisms we are studying have existed as species for 2–5 million years.

The message for ecologists studying stream fishes is clearly that these species are carrying considerable “historical baggage.” Also, it is most realistic to expect that adaptations to reduce competition or predation should be generalized rather than specific to particular species or microhabitats. Many studies of fish resource partitioning attempt to interpret interspecific differences in resource use as due solely to local spatio-temporal interactions, ignoring the evolutionary history of species. The history of the southeastern fish assemblages tells us that each species in an assemblage represents a long, spatially integrated, and potentially independent evolutionary history, and at any given point a species may not even be well adapted to its environment (Jaksic 1981).

Whether or not assemblages vary significantly over time has been a major issue in the attempts to understand factors responsible for maintaining community structure (e.g., Grossman et al. 1982; Ross

et al. 1985, 1987; Matthews et al. 1988). The available literature on stream fishes indicates that persistence of assemblages may vary between harsh and benign habitats, but that values of well over a decade are not uncommon. However, the upper limits of temporal persistence are essentially all due to the length of the study period rather than an actual limit to persistence of the assemblage (Ross et al. 1987). The actual age of assemblages (considered from a persistence viewpoint) may range from a matter of years or decades—perhaps upwards to the age of river systems, on the order of 8,000–100,000 years. For instance, the drainage characteristics of many stream systems changed significantly during the Pliocene–Pleistocene transition due to glaciation, sediment transport, withdrawal of marine transgressions, and coastal uplift (Swift et al. 1986; Mayden 1987b). Thus, many systems date from the last 10,000 years.

As a further complication, all species pairs in a contemporary assemblage may (and probably do) differ in the lengths of their associations. Rarely should we expect that an assemblage, once formed, has existed unchanged over time. For instance, Mayden (1987a) points out that we need to distinguish between vicariant and dispersive events in the formation of fish assemblages. There are probably older core assemblages, due to historical vicariant events, that have periodically gained species from dispersive events. Thus, the degree of expected coevolution should vary, even within an assemblage.

The duration of individual populations is on a much shorter time scale. Most southeastern stream fishes have fairly short lifespans of 2–7 years. For instance, red shiners generally do not live beyond 3 years (Farringer et al. 1979), darters may live 3–4 years (Page 1983), topminnows may live 4–5 years (Fisher 1981), and centrarchids may live more than 10 years (Carlander 1977). Local microhabitats also may change on a scale from hours to years, often operating on temporal scales that approach or exceed the life span of individuals within a species. The temporal duration of microhabitats varies strongly with geographical region.

The second problem is the tacit assumption that present-day conditions of seemingly undisturbed systems represent the ecosystems in which the stream fauna evolved or, at least, the conditions that the assemblage has encountered for the majority of its history. In other words, we want to be able to make generalizations from our work that rest on the assumption that ours is a natural system, essentially unchanged from prehistoric times—the fallacy of the primeval stream. Historical changes of the last 200 years (such as widespread deforestation during

the late 1800's and early 1900's) are documented well enough to argue strongly that there are virtually no southeastern streams that have escaped some degree of man-induced habitat change.

The third problem is less an outcome of our mortality than of our ability to ignore certain life history stages while emphasizing others; however, this problem is definitely related to time limitations—the fallacy of ontogenetic stasis. In a review of resource partitioning studies of fish assemblages, Ross (1986) listed only one study that examined both larval and adult life history stages. Thus, most of what we know about fish resource use in general, and certainly of stream fish resource use, is from only one or two life history stages. A quick perusal of the issues of *Current Contents* from 1986 to the present, as well as searches of *Transactions of the American Fisheries Society* and *Copeia* for 1985–88, indicates that there is less emphasis on larval ecology of freshwater fishes than on larval ecology of marine fishes. It is safe to generalize that the life cycle and associated habitat use are not completely known for a majority of southeastern stream fishes. The critical question is whether the lack of knowledge of early life history stages significantly weakens our understanding of resource requirements of the fishes and any management plans based on those requirements. Available data indicate that an understanding of resource requirements of fishes and the subsequent management plans are substantially compromised if they do not include information from early life history stages.

In summary, finite careers and budgets encourage studies of communities and stream systems on short timescales and on life history stages that are most readily collected, observed, or identified. This has led to three basic errors: (1) ignoring historical effects in the structuring of communities; (2) assuming that present day studies deal with pristine systems; and (3) focusing on the most easily studied life history stages while ignoring stages that may, in fact, be more critical to an understanding of community function.

Further understanding of assemblage formation and maintenance requires a broader scope—one that considers ongoing as well as historical effects, and one that recognizes that both biotic and abiotic forces may act differently on the various life history stages.

## Information Sources

Baker, J. A., and S. T. Ross. 1981. Spatial and temporal resource utilization by southeastern cyprinids. *Copeia* 1981:178–189.

- Carlander, K. D. 1977. Handbook of freshwater fisheries biology. Vol. 2. Iowa State University Press, Ames. 431 pp.
- Collette, B. B., and P. Banarescue. 1977. Systematics and zoogeography of the fishes of the family Percidae. *J. Fish. Res. Board Can.* 34:1450-1463.
- Farringer, R. T., III, A. A. Echelle, and S. F. Lehtinen. 1979. Reproductive cycle of the red shiner, *Notropis lutrensis*, in central Texas and south central Oklahoma. *Trans. Am. Fish. Soc.* 108:271-276.
- Fisher, J. W. 1981. Ecology of *Fundulus catenatus* in three interconnected stream orders. *Am. Midl. Nat.* 106:372-378.
- Gilbert, C. R. 1964. The American cyprinid fishes of the subgenus *Luciulus* (genus *Notropis*). *Bull. Fla. State Mus.* 8, 2. 194 pp.
- Gilbert, C. R. 1976. Composition and derivation of the North American freshwater fish fauna. *Fla. Sci.* 39:103-111.
- Grossman, G. D., P. B. Moyle, and J. O. Whittaker, Jr. 1982. Stochasticity in structural and functional characteristics of an Indiana stream fish assemblage: a test of community theory. *Am. Nat.* 120:423-454.
- Jaksic, F. M. 1981. Recognition of morphological adaptations in animals: the hypothetico-deductive method. *BioScience* 31:667-670.
- Matthews, W. J., R. C. Cashner, and F. P. Gelwick. 1988. Stability and persistence of fish faunas and assemblages in three midwestern streams. *Copeia* 1988:947-957.
- Mayden, R. L. 1987a. Historical ecology and North American highland fishes: a research program in community ecology. Pages 210-222 in W. J. Matthews and D. C. Heins, eds. Community and evolutionary ecology of North American stream fishes. University of Oklahoma Press, Norman. 310 pp.
- Mayden, R. L. 1987b. Pleistocene glaciation and historical biogeography of North American central-highland fishes. Pages 141-151 in W. C. Johnson, ed. Quaternary environments of Kansas. Kan. Geol. Surv., Guidebook Ser. 5. 208 pp.
- Page, L. M. 1983. Handbook of darters. T.F.H. Publishers, Neptune City, N.J. 271 pp.
- Ross, S. T. 1986. Resource partitioning in fish assemblages: a review of field studies. *Copeia* 1986:352-358.
- Ross, S. T., and J. A. Baker. 1983. The response of fishes to periodic spring floods in a southeastern stream. *Am. Midl. Nat.* 109:1-14.
- Ross, S. T., J. A. Baker, and K. E. Clark. 1987. Micro-habitat partitioning of southeastern stream fishes: temporal and spatial predictability. Pages 42-51 in W. J. Matthews and D. C. Heins, eds. Community and evolutionary ecology of North American stream fishes. University of Oklahoma Press, Norman.
- Ross, S. T., J. Knight, and D. Wilkins. 1989. Longitudinal occurrence of the bayou darter (Percidae: *Etheostoma rubrum*) in Bayou Pierre—a response to stream order or habitat availability? *Pol. Arch. Hydrobiol.* In press.
- Ross, S. T., W. J. Matthews, and A. A. Echelle. 1985. Persistence of stream fish assemblages: effects of environmental change. *Am. Nat.* 126:24-40.
- Swift, C. C., C. R. Gilbert, S. A. Bortone, G. H. Burgess, and R. W. Yerger. 1986. Zoogeography of the freshwater fishes of the southeastern United States: Savannah River to Lake Pontchartrain. Pages 213-265 in C. H. Hocutt and O. E. Wiley, eds. The zoogeography of North American freshwater fishes. John Wiley & Sons, New York. 866 pp.
- Swift, C. C., and E. S. Wing. 1968. Fossil bony fishes from Florida. *The Plaster Jacket*, Fla. State Mus. 7:1-11.

## A Pluralistic View of Factors Influencing Warmwater Stream Fishes: Implications for Environmental Management

by

Isaac J. Schlosser

*University of North Dakota  
Department of Biology  
Box 8238, University Station  
Grand Forks, North Dakota 58202*

Fish mortality is usually greatest during early life stages. Therefore, establishing what determines growth and mortality of fishes during early life stages will be essential for understanding fish population dynamics and for predicting the effect of environmental modifications on fishery resources. Unfortunately, our understanding of the influence of abiotic and biotic factors on early life stages of fishes in warmwater streams is poorly developed. Conducting controlled experiments is difficult and we lack a conceptual framework for identifying critical ecological processes.

In a previous descriptive study (Schlosser 1982a), I examined temporal and spatial variation in fish community structure along a pronounced habitat gradient. Based on these results I proposed a conceptual framework that attempted to integrate the relative effect of abiotic and biotic factors on fishes in small warmwater streams (Schlosser 1987a). From this conceptual framework it was evident that three factors potentially have major effects on growth and survival of early life stages of stream fishes: (1) harsh winter conditions, (2) direct effects of flow regime on invertebrate resource availability and fish survival, and (3) trophic interactions, including predation and competition.

My recent research used both descriptive and experimental approaches to examine the effects of trophic interactions and flow regimes on juvenile fishes. Results of this research indicated that (1) spring floods during spawning periods are associated with a major decrease in the density of juvenile cyprinid and centrarchid fishes (Schlosser 1985; see also Harvey 1987; Bain et al. 1988); (2) elevated (nonscouring) flow results in a dramatic increase in invertebrate abundance during spring and summer, when most growth of juvenile fishes is likely to occur (Schlosser and Ebel 1989); (3) in the absence of predators, small fishes depress invertebrate abundance

in pools but not in riffles (Schlosser and Ebel 1989); and (4) centrarchid predators restrict juvenile fishes to shallow refugia while engaging in both species- and size-selective predation (Schlosser 1987b, 1988b).

These results indicate that multiple abiotic (e.g., channel morphology, flow regime) and biotic (e.g., resource depression, predation) factors interact to influence growth and survival of early life stages of stream fishes. Because many environmental modifications simultaneously alter several of these variables, such modifications are likely to have considerable effects on the fishery resources in modified streams. For example, agricultural land use alters flow regime, channel morphology, seasonal timing of resource availability, and abundance of piscivorous fishes (Karr and Schlosser 1978; Schlosser 1982b). These modifications, in turn, have major effects on temporal variability and juvenile recruitment in headwater and downstream areas (Schlosser 1982b).

These results also indicate that the development of a predictive paradigm for environmental and fisheries management in streams will require refinement in our understanding of how abiotic and biotic factors interact to affect early life stages of stream fishes. This refinement will be achieved through a combination of (1) long-term monitoring to establish the influence of natural environmental variability on growth and survival of early life stages and (2) experimental studies to assess the effect of variability in flow and temperature regime on trophic interactions. Lastly, fishes are highly migratory during reproductive activities, and stream systems are heterogeneous and interconnected. Hence, it will be necessary to explore the effects environmental modification in one stream have on population and community dynamics of fishes in adjacent parts of the stream system (landscape approach of Forman and Godron 1986).

## Information Sources

- Bain, M. B., J. T. Finn, and H. E. Booke. 1988. Streamflow regulation and fish community structure. *Ecology* 69:382-392.
- Forman, R. T. T., and M. Godron. 1986. *Landscape ecology*. John Wiley & Sons, New York. 619 pp.
- Harvey, B. C. 1987. Susceptibility of young-of-the-year fishes to downstream displacement by flooding. *Trans. Am. Fish. Soc.* 116:851-855.
- Karr, J. R., and I. J. Schlosser. 1978. Water resources and the land-water interface. *Science* 201:229-234.
- Schlosser, I. J. 1982a. Fish community structure and function along two habitat gradients in a headwater stream. *Ecol. Monogr.* 52:395-414.
- Schlosser, I. J. 1982b. Trophic structure, reproductive success, and growth rate of fishes in a natural and modified headwater stream. *Can. J. Fish. Aquat. Sci.* 39:968-978.
- Schlosser, I. J. 1985. Flow regime, juvenile abundance, and assemblage structure of stream fishes. *Ecology* 66:1484-1490.
- Schlosser, I. J. 1987a. A conceptual framework for fish communities in small warmwater streams. Pages 17-24 in W. J. Matthews and D. C. Heins, eds. *The ecology and evolution of North American stream fish communities*. University of Oklahoma Press, Norman. 310 pp.
- Schlosser, I. J. 1987b. The role of predation in age- and size-related habitat use by stream fishes. *Ecology* 68:651-659.
- Schlosser, I. J. 1988a. Predation rates and the behavioral response of adult brassy minnows (*Hybognathus hankinsoni*) to creek chub and smallmouth bass predators. *Copeia* 1988:691-697.
- Schlosser, I. J. 1988b. Predation risk and habitat selection by two size classes of a stream cyprinid: experimental test of a hypothesis. *Oikos* 52:36-40.
- Schlosser, I. J., and K. K. Ebel. 1989. Effects of flow regime and cyprinid predation on a headwater stream. *Ecol. Monogr.* 59:41-57.

## Fish Community Structure and Stability in Warmwater Midwestern Streams

by

William J. Matthews

*University of Oklahoma  
Biological Station  
Kingston, Oklahoma 73439*

Impact assessment generally implies a search for ways to measure and predict changes in stream systems that ensue from cultural alterations. In order to really know the degree to which a cultural activity results in change in stream systems, something must be known of the magnitude of change or variation that occurs naturally. Even if managers are constrained from long-term studies of each stream of interest, the long-term data sets that do exist can be used to predict the kinds and degree of changes that may naturally occur in a variety of stream systems.

Over the last two decades, a relatively long-term data base has been accumulated from our collecting activities assessing fish assemblages and faunas in three midwestern streams: Piney Creek, Izard County, Arkansas; Brier Creek, Marshall County, Oklahoma; and Kiamichi River, southeastern Oklahoma (Ross et al. 1985; Matthews 1986; Matthews et al. 1988). These data sets have been evaluated at two levels: stability of whole-stream faunas (based on pooled collection data at multiple sites on each stream) and stability of fish assemblages at each individual site over time. The data set spans 1969–86 for Brier Creek, 1972–86 for Piney Creek, and 1981–86 for Kiamichi River. Each of these time intervals allows for one to two complete generations of at least the small fish species that are most abundant.

The whole-stream faunas of all three streams have been highly persistent (in terms of presence and absence of species) and stable (based on abundance data) in all years of study. No common species has been lost from any of the streams, and no “rare” species has become common at any of the sites. Both similarity indices (Morisita’s and percent similarity) and statistical rank correlation of species abundances show that all three streams have been stable, and that the most environmentally stable stream (Piney Creek of the Ozark uplands) has had the

greatest degree of fish population stability across all years.

At individual sites on each of the streams, the local fish assemblage has had a tendency to be stable, although exceptions exist in all three systems. Where individual collecting sites have shown a marked change in the composition of the fish assemblage, this change can usually be attributed to known events, such as deposition of large woody debris by floods, death of fish in intermittent headwaters due to drought, or other such disturbances. However, recolonization of intermittent headwaters is not a random phenomenon. Matthews (1987) showed that existence and persistence of fish species in harsh headwaters of at least one of the streams was directly related to their ability to withstand low oxygen conditions, which are common. Thus, there is both a chance component and a directed component to dynamics of assemblage composition in headwater streams.

The long-term study of Piney Creek included a flood event that was clearly of 100-year magnitude. Vertical water levels were elevated as much as 12 m at some sites, and physical destruction and disturbance of habitat along the creek was extreme. Immediately after the flood, fish assemblages showed some differences from those that existed before the flood, but after 8 months, both the whole-stream fauna and most local assemblages were statistically indistinguishable from their composition before the flood. This well-documented event—and other accounts of flood effects on fish in midwestern streams (Gelwick 1990; Harvey 1987)—indicates that while floods may have an immediate effect on composition of midwestern fish communities, these changes are largely transient, with the systems returning to pre-flood conditions rather rapidly.

Some methodological recommendations are possible from collections made on these streams. There is a longstanding question in fish community assess-

ment as to what length of stream constitutes a most appropriate and cost-effective collecting activity. In many standardized procedures, a reach of 100 m has been adopted for small- to medium-sized streams. In contrast, my collections at a site have always included a minimum of 200 m—often up to 400 m—of stream, in order to sample sufficient distance to find representative habitat for all species that are present. In 1988, I compared fish collections in adjacent 100-m sections of Brier Creek at a total of 5 sites, or 10 segments. Typically, the overall fish assemblage in spatially adjacent 100-m segments had percent similarity in composition of only 60–70%, and hence a different picture of the fish community resulted if a total of 200 m was included in the sample instead of only 100 m. Accordingly, in at least some midwestern streams, I suggest that more than 100 m of stream reach should be sampled, whether the goal of the study is assessment of some particular effect or the acquisition of long-term data on basic ecological properties of the fish community.

## Information Sources

- Gelwick, F. P. 1990. Longitudinal and temporal comparisons of riffle and pool fish assemblages in an Ozark stream. *Copeia* 1990. In press.
- Harvey, B. C. 1987. Susceptibility of young-of-the-year fishes to downstream displacement by flooding. *Trans. Am. Fish. Soc.* 116:851–855.
- Matthews, W. J. 1986. Fish faunal structure in an Ozark stream: stability, persistence, and a catastrophic flood. *Copeia* 1986:388–397.
- Matthews, W. J. 1987. Physicochemical tolerance and selectivity of stream fishes as related to their geographic ranges and local distributions. Pages 111–120 in W. J. Matthews and D. C. Heins, eds. *Community and evolutionary ecology of North American stream fishes*. University of Oklahoma Press, Norman. 310 pp.
- Matthews, W. J., R. C. Cashner, and F. P. Gelwick. 1988. Stability and persistence of fish faunas and assemblages in three midwestern streams. *Copeia* 1988:947–957.
- Ross, S. T., W. J. Matthews, and A. A. Echelle. 1985. Persistence of stream fish assemblages: effects of environmental change. *Am. Nat.* 126:24–40.

## Centrarchid-Habitat Associations in Ozark Streams

by

Charles F. Rabeni

*U.S. Fish and Wildlife Service  
Missouri Cooperative Fish and Wildlife Research Unit  
112 Stephens Hall, University of Missouri  
Columbia, Missouri 65211*

Any goal of sportfish management should include satisfactory sustained angling benefits, which can occur only by maintaining satisfactory habitat and balanced fish populations and communities. Appropriate habitat is an important prerequisite for balanced fish populations in streams. Yet, not enough is known about habitat requirements of most fish. Unless the stream is highly degraded, it is not possible to take a generalized habitat model developed from the literature and apply it to a particular stream with much hope of identifying ways to increase fishing quality. Each stream, or at least each stream type, must be considered individually because fish are often plastic in their behavior and requirements, and a limiting variable in one geographic area may be unimportant in another.

Two approaches have been taken by researchers working on species-habitat relations. The first is to determine how individual fish use particular habitat elements and then infer something about the entire population. This is dangerous because there is no known empirical linkage between the two. Drawing ecological meaning from data on habitat use or preference is similar to trying to evaluate mechanisms from correlations. Habitats of a stream fish are often measured in terms of either absolute or relative use, yet these data reveal nothing about how important a particular habitat variable might be nor how the variables are interrelated.

A second approach is to correlate some population characteristics, generally density or biomass, with some habitat feature. While some studies are successful in relating fish quantity to some habitat quantity, their usefulness in a management context is limited. There is generally no consistent relation between fish amounts and habitat amounts. Although a biologist can often distinguish a "good" fishing stream from a "bad" one, unless a stream is initially severely degraded there is little evidence that altering a particular habitat variable will cause

a predictable corresponding change in the fish population. We have little understanding of factors affecting carrying capacity of a warmwater stream—especially the relation between food and habitat. While our goal is to quantify critical factors of the stream environment, it does not appear that this goal can be attained in the near future. What we as fishery researchers should do is isolate a subset of environmental variables, pertinent to the particular situation at hand, from a larger set of variables considered generally important to a species—such as those listed in the U.S. Fish and Wildlife Service's Habitat Suitability Index Models. If done carefully, we should be able to pinpoint particularly important habitat variables that influence the success of a population and on which management efforts can be focused.

Habitat conditions can be related to a life stage in one of three ways. Nonuseful habitat conditions are those not required for the population's survival and may have an adverse relation to a population (e.g., extremely high water velocity). Useful habitat conditions provide a necessary element for the population's well-being when other habitats also provide the same elements (e.g., rootwads and log jams). Essential habitats are those required for the population's well-being.

We must determine which habitat elements fit into which category to effectively manage a species. Unfortunately, nonuseful habitats are often impossible to identify because even if an area is unused by fish it may still be important as a food-source area or barrier to predators or competitors. Useful habitats can be detailed if the species is studied in more than one location. Essential habitat features may or may not be limiting to a population.

Based on the preceding concepts, I evaluated the habitat relations of Ozark stream centrarchids, with emphasis on smallmouth bass (*Micropterus dolomieu*) and lesser emphasis on rock bass (*Ambloplites*

*rupestris*) and longear sunfish (*Lepomis megalotis*). I studied behavior of individual fish by underwater observation and telemetry; documented population characteristics of density, biomass, and condition; and attempted to relate habitat use by individuals to the well-being of the population.

I documented daytime habitat use in summer using underwater observation. After I corrected for differences in the amount of open water and each cover type observed, over 90% of individuals of each species were found to be positioned within 1 m of cover. Overlaps in cover use between species suggested that cover is not important in interspecific segregation. Species segregation was strongest by current velocity, with smallmouth bass using cover in the fastest water and longear sunfish using cover in the slowest water. Current velocity was also important to smallmouth bass and longear sunfish as an element of intraspecific segregation. Strong positive linear relations between water depth and fish size were noted for all three species, but water depth was not important in interspecific segregation. Substrate use appeared to be dependent on cover use, and results were inconclusive. All three species shifted habitat throughout the day, but the greatest shifts occurred at night.

Telemetry studies on smallmouth bass substantiated the observational studies and added information on fish habitat associations in winter, when fish used boulders almost exclusively. Both studies showed that smallmouth bass avoided shallow areas and existed almost exclusively in current velocities  $<0.2$  m/s. I believe that the results of studies emphasizing individual fish can be used with other studies to delineate essential habitat conditions for smallmouth bass populations.

Regression models from population studies showed that depth and current did not correlate with standing crop of smallmouth bass. Perhaps these variables are important but are available in greater amounts than needed. What the regression model did show is that boulders had a positive relation

to standing crops. Perhaps boulders are an essential habitat or a limiting factor for smallmouth bass in these streams because of some important, yet still unclear, function during cold-water periods. Fish may be more energetically efficient when associated with boulders during stressful periods. The significant, positive, but weak relation between boulders and fish density or biomass indicates that fish use boulder areas in a nonlinear fashion. Underwater observations indicated that a single boulder properly situated could accommodate as many as eight adult smallmouth bass. Thus, while a particular aspect of the habitat may be essential, its quality may be far more important than its quantity. These studies suggest that obtaining the information necessary for effective management of a fish population requires several approaches, and all these approaches have to be repeated for different life stages.

## Information Sources

- Covington, W. G., R. E. Marteney, and C. F. Rabeni. 1983. Population characteristics of rock bass and smallmouth bass in the Ozark National Scenic Riverways. *Trans. Mo. Acad. Sci.* 17:27-35.
- McClendon, D. D., and C. F. Rabeni. 1986. A comparison of electrofishing and underwater observations in evaluating population characteristics of stream centrarchids. *Proc. Annu. Conf. Southeast. Assoc. Fish Wildl. Agencies* 40:92-101.
- Probst, W., C. F. Rabeni, C. G. Covington, and R. E. Marteney. 1984. Resource partitioning between stream-dwelling rock bass (*Ambloplites rupestris*) and smallmouth bass (*Micropterus dolomieu*). *Trans. Am. Fish. Soc.* 113:283-294.
- Rabeni, C. F., D. D. McClendon, W. Probst, B. Todd, A. Livingstone, and D. I. Foster. 1990. Management of smallmouth bass (*Micropterus dolomieu*) in the Ozark National Scenic Riverways. Proceedings of the Conference on Science in the National Parks, July 1986, Fort Collins, Colo. In press.
- Todd, B. L., and C. F. Rabeni. 1989. Movement and habitat use by stream-dwelling smallmouth bass. *Trans. Am. Fish. Soc.* 118:229-242.

## Fish Larvae—Ecologically Distinct Organisms<sup>1</sup>

by

Darrel E. Snyder

Larval Fish Laboratory  
Colorado State University  
Fort Collins, Colorado 80523

The behaviors and habitat requirements of larvae of most warmwater stream fishes are distinctly different from those of their juvenile and adult counterparts (Braum 1978; Marcy et al. 1980; Snyder 1983, 1985; Floyd et al. 1984; Faber 1985). The larval period—the interval of fish development from hatching or birth to loss of finfolds and development of adult complement of fin spines and rays—entails many and often dramatic changes in morphology and physiology. Also, it usually covers a number of short-term shifts in lifestyle and habitat. Indeed, a fish larva is often ecologically distinct not only from its juvenile and adult counterparts but from itself during certain intervals within the larval period.

The initial habitat of most fish larvae is the spawning ground. For warmwater stream fishes and anadromous species, there is much diversity in the earliest habitat requirements and behavior of larvae. Initial larval habitats, associated environmental requirements, and behaviors are considered in a classification of reproductive guilds by Balon (1981, 1984). The 22 families and over 260 species (Swift et al. 1986) of freshwater and anadromous fishes in the southeastern United States represent at least 17 of Balon's reproductive guilds (Table).

Some fishes remain on or near the spawning grounds throughout much or all of their early development, and some may use the same habitat throughout their life cycle. However, for most stream fishes, the initial habitat rapidly becomes inadequate, and the larvae either drift or actively migrate to more suitable nursery grounds, sometimes hundreds of kilometers downstream. The larvae and early juveniles of most stream fishes tend to use nearshore areas with relatively slow-moving water near cover, vegetation, or sharp vertical relief.

Larval fish size largely dictates the characteristics of suitable nursery habitat and the role larvae play in the aquatic system. In southeastern fresh waters, fish larvae may be as small as 2 mm in total length (TL) at hatching (e.g., white crappie [*Pomoxis anularis*]) and early larvae of some (e.g., striped bass [*Morone saxatilis*], freshwater drum [*Aplodinotus grunniens*], and emerald shiner [*Notropis atherinoides*]) inhabit pelagic waters and constitute a part of the planktonic community. These early pelagic larvae may be food for predatory copepods, whereas later larvae of these fishes may reverse the roles and prey on the copepods. Most fish larvae feed on rotifers and other small zooplankton; however, some are piscivorous and may consume other fish larvae nearly as long as themselves. Clark and Pearson (1979) observed fish larvae in the stomachs of over 25% of small (4–5 mm standard length [SL]) freshwater drum larvae. For this species, piscivory decreased with size and was not observed in larvae or early juveniles over 9 mm SL.

For most stream fishes, extremely high mortality during the embryonic and larval period is normal and accommodated by their reproductive strategy. Environmental effects that substantially add to, or reduce, natural larval fish mortality can have a corresponding effect on the eventual size of the adult population. The effects of changes in habitat and community structure on fish larvae can be quite different from those on juveniles and adults. The most common effect is loss or alteration of larval fish habitat. Fish larvae can be particularly sensitive to physical and chemical water pollution. Biotic alterations that might not be expected to have an adverse effect on native fishes, such as the introduction of exotic forage species, might indeed affect native populations by excess predation on, or competition with, their larvae. Fish larvae of many species are especially vulnerable to entrainment in water withdrawal systems for irrigation, domestic and industrial water supplies, and power plant cooling.

<sup>1</sup>Contribution 42 of the Larval Fish Laboratory, Colorado State University.

Table. *Reproductive guilds<sup>a</sup> by family for fishes in fresh waters of the southeastern United States.*

| Family                                  | Number<br>of<br>guilds | Nonguarders    |                |                |     |     |     | Guarders        |                            |         |     | Bearers |               |          |   |      |   |   |
|---|------------------------|----------------|----------------|----------------|-----|-----|-----|-----------------|----------------------------|---------|-----|---------|---------------|----------|---|------|---|---|
|   |                        | Open substrate |                |                |     |     |     | Brood<br>hiders | Sub-<br>strate<br>choosers | Nesters |     |         | Exter-<br>nal | Internal |   |      |   |   |
|   |                        | A11            | A12            | A13            | A14 | A15 | A16 |                 |                            | B22     | B23 | B25     | B27           |          |   |      |   |   |
| Acipenseridae<br>(sturgeons)            | 2                      |                | X <sup>b</sup> | X              |     |     |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Polyodontidae<br>(paddlefish)           | 1                      |                |                | X <sup>b</sup> |     |     |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Lepisosteidae<br>(gars)                 | 1                      |                |                |                |     |     | X   |                 |                            |         |     |         |               |          |   |      |   |   |
| Amiidae<br>(bowfin)                     | 1                      |                |                |                |     |     |     |                 |                            |         |     |         |               |          |   | X    |   |   |
| Clupeidae<br>(herrings)                 | 3                      | X              | X              |                |     | X   |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Hiodontidae<br>(mooneyes)               | 1                      |                |                | X              |     |     |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Salmonidae<br>(trouts)                  | 1                      |                |                |                |     |     |     | X               |                            |         |     |         |               |          |   |      |   |   |
| Umbridae<br>(mudminnows)                | 1                      |                |                |                |     |     |     |                 |                            |         |     |         |               | X        |   |      |   |   |
| Esocidae<br>(pikes)                     | 1                      |                |                |                |     | X   |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Cyprinidae<br>(minnows)                 | 9                      | X              |                | X              | X   | X   | X   | X               | X                          |         |     |         |               | X        |   | X    |   |   |
| Catostomidae<br>(suckers)               | 3                      |                |                | X              |     |     | X   | X               |                            |         |     |         |               |          |   |      |   |   |
| Ictaluridae<br>(catfishes)              | 2                      |                |                |                |     |     |     |                 |                            |         |     |         |               | X        |   | X    |   |   |
| Amblyopsidae<br>(cavefishes)            | 1                      |                |                |                |     |     |     |                 |                            |         |     |         |               |          |   | X    |   |   |
| Aphredoderidae<br>(pirate perch)        | 1                      |                |                |                |     |     |     |                 |                            |         |     |         |               | ?        |   | ?    |   |   |
| Cyprinodontidae<br>(killifishes)        | 3                      |                |                |                | X   | X   |     |                 |                            |         |     |         |               | X        |   |      |   |   |
| Poeciliidae<br>(livebearers)            | 2                      |                |                |                |     |     |     |                 |                            |         |     |         |               |          |   | X    | X |   |
| Atherinidae<br>(silversides)            | 1                      |                |                |                | X   |     |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Percichthyidae<br>(temperate<br>basses) | 2                      | X              |                |                |     | X   |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Centrarchidae<br>(sunfishes)            | 4                      |                |                |                |     |     |     |                 |                            | X       |     | X       | X             | X        |   |      |   |   |
| Percidae<br>(perches)                   | 7                      |                | X              |                | X   | X   | X   | X               | X                          |         |     |         |               | X        | X |      |   |   |
| Sciaenidae<br>(drum)                    | 1                      | X              |                |                |     |     |     |                 |                            |         |     |         |               |          |   |      |   |   |
| Cottidae<br>(sculpins)                  | 1                      |                |                |                |     |     |     |                 |                            |         |     |         |               | X        |   |      |   |   |
| Number of<br>families                   |                        | 4              | 5              | 3              | 6   | 6   | 3   | 3               | 1                          | 1       | 1   | 1(2)    | 3             | 4        | 4 | 1(2) | 1 | 1 |

<sup>a</sup>Guild codes: A11 = pelagic spawners; A12 = rock and gravel spawners with pelagic larvae; A13 = rock and gravel spawners with benthic larvae; A14 = nonobligatory plant spawners; A15 = obligatory plant spawners; A16 = sand spawner; A23 = rock and gravel hiders; A24 = cavity hiders; B13 = rock and gravel tenders; B14 = plant tenders; B22 = miscellaneous substrate nesters; B23 = rock and gravel nesters; B25 = plant material nesters; B27 = hole nesters; C14 = gill-chamber brooders; C22 = obligate lecithotrophic livebearers; C24 = viviparous trophoderms (see Balon 1975, 1981, 1984 for detailed descriptions of these and other guilds).

<sup>b</sup>Between A12 and A13—recently hatched larvae drift near bottom (epibenthic drifters?).

In our attempt to evaluate environmental impacts and aquatic management programs, we often turn to models, habitat-quality indices, and related tools. Unfortunately, these models or tools can be no better than the data on which they are based, and for most species of fish, reproductive and early life history data are often lacking, inadequate, or questionable. For southeastern fishes and aquatic systems, there is much basic research that needs to be done to fill the immense void in our knowledge, including such basic tasks as determining what the larvae of many fishes look like. In the Southeast, we have only four limited manuals for larval fish identification (Hogue et al. 1976; McGowan 1984; Conrow and Zale 1985), although some species are covered in taxonomic manuals for other parts of the country (Auer 1982; see lists in Snyder 1983 or Simon 1986 for others). The importance of accurate identification of specimens cannot be overemphasized because critical resource management decisions are sometimes based on species-specific field data. Before we can effectively proceed with field studies on fish larvae in the Southeast, we must be able to accurately identify specimens.

The early life stages of fishes must be an important concern in the development of management plans and impact assessment methods. Aquatic ecologists and fishery biologists are often too preoccupied with the needs of adult fishes to recognize the differing requirements of earlier life stages. It is simpler and less costly to concentrate on one life stage of a target species than on a whole series of ecologically distinct stages; however, fish populations depend on adequate survival of their embryos, larvae, and early juveniles.

## Information Sources

- Auer, N. A. 1982. Identification of larval fishes of the Great Lakes Basin with emphasis on the Lake Michigan drainage. Great Lakes Fishery Commission, Ann Arbor, Mich. Spec. Publ. 82-3. 744 pp.
- Balon, E. K. 1975. Reproductive guilds of fishes: a proposal and definition. *J. Fish. Res. Board Can.* 32:821-864.
- Balon, E. K. 1981. Additions and amendments to the classification of reproductive styles in fishes. *Environ. Biol. Fishes* 6:377-389.
- Balon, E. K. 1984. Patterns in the evolution of reproductive styles in fishes. Pages 35-53 in G. W. Potts and R. J. Wootton, eds. *Fish reproduction, strategies and tactics*. Academic Press, Inc., New York. 410 pp.
- Balon, E. K. 1985. The theory of saltatory ontogeny and life history models revisited. Pages 13-30 in E. K. Balon, ed. *Early life histories of fishes; new developmental, ecological and evolutionary perspectives*. W. Junk Publishers, Dordrecht, The Netherlands. 280 pp.
- Balon, E. K., W. T. Momot, and H. A. Regier. 1977. Reproductive guilds of percids: results of the paleogeographic history and ecological succession. *J. Fish. Res. Board Can.* 34:1910-1921.
- Bennett, D. H., and R. W. McFarlane. 1983. The fishes of the Savannah River Plant: National Environmental Park. U.S. Department of Energy, National Environmental Research Park, Savannah River Ecology Laboratory, Aiken, S.C. 152 pp.
- Blaxter, J. H. S. 1969. Development: eggs and larvae. Pages 177-252 in W. S. Hoar and D. J. Randall, eds. *Fish physiology. Volume 3: reproduction and growth, bioluminescence, pigments and poisons*. Academic Press, Inc., New York. 485 pp.
- Blaxter, J. H. S. 1986. Development of sense organs and behavior of teleost larvae with special reference to feeding and predator avoidance. *Trans. Am. Fish. Soc.* 115:98-114.
- Braum, E. 1978. Ecological aspects of the survival of fish eggs, embryos and larvae. Pages 102-134 in S. D. Gerking, ed. *Ecology of freshwater fish production*. John Wiley & Sons, New York. 520 pp.
- Clark, A. L., and W. D. Pearson. 1979. Early piscivory in larvae of the freshwater drum, *Aplodinotus grunniens*. Pages 31-59 in R. Wallus and C. W. Voigtlander, eds. *Proceedings of a workshop on freshwater larval fishes*. Tennessee Valley Authority, Norris, Tenn. 241 pp. [There are some size-related and species identification errors in this paper, including the smallest specimen drawn and photographed: the specimen shown in Figs. 3 and 7, labeled a 3.4-mm freshwater drum, was actually a percid, probably *Stizostedion*, at least 8 mm.]
- Conrow, R., and A. V. Zale. 1985. Early life history stages of fishes of Orange Lake, Florida: an illustrated identification manual. Fla. Coop. Fish Wildl. Res. Unit Tech. Rep. 15, University of Florida, Gainesville. 45 pp.
- Faber, D. J. 1985. Water babies: larval fishes of Ottawa and vicinity. National Museum of Natural Sciences, Ottawa, Ontario. 25 pp.
- Floyd, K. B., R. D. Hoyt, and S. Timbrook. 1984. Chronology of appearance and habitat partitioning by stream larval fishes. *Trans. Am. Fish. Soc.* 113:217-223.
- Hardy, J. D., Jr., G. E. Drewry, R. A. Fritzsche, G. D. Johnson, P. W. Jones, and F. D. Martin. 1978. Development of fishes of the Mid-Atlantic Bight, an atlas of egg, larval and juvenile stages. U.S. Fish. Wildl. Serv., FWS/OBS-78/12. 6 volumes.
- Hoar, W. S., and D. J. Randall, editors. 1988. *Fish physiology. Volume 11: the physiology of developing fish*. Academic Press, Inc., New York. Part A, 546 pp.; Part B, 436 pp.
- Hogue, J. J., Jr., R. Wallus, and L. K. Kay. 1976. Preliminary guide to the identification of larval fishes in the Tennessee River. Tennessee Valley Authority, Technical Note B19, Knoxville, Tenn. 66 pp.
- Holland, L. E., and M. L. Huston. 1983. A compilation of available literature on the larvae of fishes common to the upper Mississippi River. U.S. Fish and Wildlife Service, National Fishery Research Laboratory, La Crosse, Wis. 364 pp.
- Kindschi, G. A., R. D. Hoyt, and G. J. Overmann. 1979. Some aspects of the ecology of larval fishes in Rough River Lake, Kentucky. Pages 139-166 in R. D. Hoyt, ed. *Proceedings of the third symposium on larval fish*. Western Kentucky University, Department of Biology, Bowling Green. 236 pp.

- Laerm, J., and B. J. Freeman. 1986. Fishes of the Okefenokee Swamp. University of Georgia Press, Athens. 118 pp.
- Lathrop, B. F., and D. E. Snyder. 1986. Bibliographies on fish reproduction and early life history. Am. Fish. Soc. Early Life Hist. Sect. Newsl. 7(2):29–36.
- Marcy, B. C., V. R. Kranz, and R. P. Barr. 1980. Ecological and behavioral characteristics of fish eggs and young influencing their entrainment. Pages 29–74 in C. H. Hocutt, J. R. Stauffer, Jr., J. E. Edinger, L. W. Hall, Jr., and R. P. Morgan, II, eds. Power plants: effects on fish and shellfish behavior. Academic Press, Inc., New York. 346 pp.
- McGowan, E. G. 1984. An identification guide for selected larval fishes from Robinson Impoundment, South Carolina. Carolina Power and Light Company, New Hill, N.C. 56 pp.
- McGowan, E. G. 1988. An illustrated guide to larval fishes from three North Carolina piedmont impoundments. Carolina Power and Light Company, New Hill, N.C. 113 pp.
- Moser, H. G., W. J. Richards, D. M. Cohen, M. P. Fahay, A. W. Kendall, Jr., and S. L. Richardson. 1984. Ontogeny and systematics of fishes. Am. Soc. Ichthyol. Herpetol. Spec. Publ. 1. 760 pp.
- Muth, R. T., and J. C. Schmulbach. 1984. Downstream transport of fish larvae in a shallow prairie river. Trans. Am. Fish. Soc. 113:224–230.
- Scott, W. B., and E. J. Crossman. 1973. Freshwater fishes of Canada. Fish. Res. Board Can. Bull. 184. 966 pp.
- Simon, T. P. 1986. A listing of regional guides, keys, and selected comparative descriptions of freshwater and marine larval fishes. Am. Fish. Soc. Early Life Hist. Sect. Newsl. 7(1):10–15.
- Snyder, D. E. 1976. Terminologies for intervals of larval fish development. Pages 41–60 in J. Boreman, ed. Great Lakes fish egg and larvae identification; proceedings of a workshop. U.S. Fish Wildl. Serv., FWS/OBS-76/23. 220 pp.
- Snyder, D. E. 1981. Contributions to a guide to the cypriniform fish larvae of the Upper Colorado River system in Colorado. U.S. Bur. Land Manage., Biol. Sci. Ser. 3. 81 pp.
- Snyder, D. E. 1983. Fish eggs and larvae. Pages 165–197 in L. Nielsen and D. Johnson, eds. Fishery techniques. American Fisheries Society, Bethesda, Md. 458 pp.
- Snyder, D. E. 1985. Larval fish ecology: a critical management concern. U.S. Fish and Wildlife Service, Office of Information Transfer, Fort Collins, Colo. [videotape, 24 min.]
- Swift, C. C., C. R. Gilbert, S. A. Bortone, G. H. Burgess, and R. W. Yerger. 1986. Zoogeography of the freshwater fishes of the southeastern United States: Savannah River to Lake Pontchartrain. Pages 213–265 in C. H. Hocutt and E. O. Wiley, eds. The zoogeography of North American freshwater fishes. John Wiley & Sons, New York. 866 pp.
- Taber, C. A. 1969. The distribution and identification of larval fishes in the Buncombe Creek Arm of Lake Texoma with observations on spawning habits and relative abundance. Ph.D. dissertation, University of Oklahoma, Norman. 120 pp.
- Wang, J. C. S., and R. J. Kernehan. 1979. Fishes of the Delaware estuaries, a guide to the early life histories. Ecological Analysts, Inc., Towson, Md. 410 pp.

# Assessment and Applied Research

The presentations by biologists involved in developing or using stream assessment methods reflect the challenges and tradeoffs of simplifying complex systems for use in practical but valid methods. The speakers of this group vary considerably in their approaches, although all are primarily interested in stream fishes. One group of speakers emphasizes the need to develop and use methods based on measures of fish communities and natural flow regimes. These biologists believe that assessment methods need to use information on all fishes rather than a few target species, and they address patterns of streamflow rather than simply minimum flow. Other speakers emphasize the practical aspects of stream assessment and the usefulness of assessment methods based on easily estimated and narrowly focused measures. These biologists deal with stream system complexity and practical constraints by advocating the use of methods that vary from simple statistical standards (e.g., median August flow, percent of mean annual flow) to computer modeling. Ironically, when the management outcomes of different assessment methods were compared (see Orth and Leonard), the differences were not that striking. The solution to appropriately simplifying the complexities of stream communities seems to be far away but may emerge from the choice of methods made by management biologists as they face future stream impact issues.

## The IBI: A Quantitative, Easily Communicated Assessment of the Health and Complexity of Entire Fish Communities<sup>1</sup>

by

Robert M. Hughes

*U.S. Environmental Protection Agency  
Environmental Research Laboratory  
NSI Technology Services Corporation  
200 SW 35th Street  
Corvallis, Oregon 97333*

Two basic questions face biologists interested in assessing and predicting impacts on stream ecosystems: Why do we need assessments of biological communities? How can data on biological communities (species and abundances) be made useful for nonbiologists (decision-makers and the public)?

Answers to these questions are central to implementing the Clean Water Act. Although the act requires restoration and maintenance of biological integrity (Section 101) and biological assessment (Sections 105, 303, 304, 305, 404), *biological integrity* remains poorly defined and rarely measured. States are required to establish water quality standards consisting of designated biological uses (broad goals) and criteria (which if met are presumed to protect the uses); however, both the uses and the national chemical criteria are inaccurate. Uses such as *aquatic life* and *warmwater fish* are so broad as to be met by any form of aquatic life or species of fish, in any abundance.

Nationally established chemical criteria ignore naturally occurring differences in conventional water quality parameters, such as dissolved oxygen, pH, and the ionic character of waters. The ionic character of waters is particularly important because it affects the activity of toxic chemicals. Equally important, single-chemical criteria do not consider combined-chemical effects, which may mitigate or magnify results. Single-species toxicity tests may not be appropriate or sensitive measures of ecosystem effects because test species may not

be representative of indigenous species or the most sensitive species and because populations can be devastated by indirect effects on competitors, predators, and prey. Furthermore, criteria exist for only a small number of chemicals. It is too expensive to develop specific criteria for every site, and it would create immense regulatory problems if they were developed. Water quality monitoring data are difficult to interpret because of the lack of an appropriate sampling and reporting framework and because of the weak link between water quality and ecosystem structure and function. Finally, there are many other factors besides toxics that may limit attainment of biological integrity.

Like toxics, physical habitat conditions are also surrogate measures of the communities that agencies are mandated to protect. Although physical and chemical conditions are necessary for understanding and explaining biological conditions, biological monitoring is preferable in many cases because it provides a direct assessment of the biological community. Moreover, the biota reflect the integrated chemical and physical quality of an area, and they often can be assessed at lower cost than physico-chemical habitat parameters, such as priority pollutants and species-specific habitat. Millions of dollars have been spent on chemical monitoring and on the development of instream flow models; still, there is only limited quantitative information regarding the health of the resident biota at those sites. In the southeastern United States (unlike the West, where the methodology developed), meeting instream flow requirements for game fish or a small number of nongame species is underprotective of the entire fish assemblage because (1) most game species have wider habitat-flow tolerances (e.g., centrarchids vs. salmonids), (2) many nongame species have narrower habitat tolerances than the game species,

<sup>1</sup>This paper was funded by the U.S. Environmental Protection Agency through Contract 68-C8-0006 to NSI Technology Services Corporation. It has been subjected to EPA's peer and administrative review and approved for publication. An earlier draft benefited from reviews by M. Bain, J. Kurtenbach, I. Schlosser, and D. Vanna-Miller.

and (3) the fish fauna has far greater species richness than in the West.

Direct biological monitoring of reference sites and sites of interest, conducted within an appropriately scaled ecoregion framework, can provide both an assessment of status and a prediction of effects, given similar stressors in similar ecoregions (Hughes et al. 1986; Omernik 1987; Hughes and Larsen 1988). Such information is generally more convincing to the public and to judges than are violations of chemical criteria and flow standards (E. Bender, USEPA Office of Water Enforcement and Permits, Washington, DC, personal communication). To increase the accuracy of our predictions, we must combine long-term biological monitoring and experimental field studies of the effects of key physical and chemical variables on populations and communities.

How can data on species and their abundances be made understandable to the public and to water resource regulators while retaining the ecological information that is meaningful to biologists? A recent attempt to communicate such information is the Index of Biotic Integrity, or IBI (Karr 1981; Karr et al. 1986). The IBI is a means of quantifying ichthyologists' judgments of the relative quality of a fish assemblage. It is based on a sample of the entire fish assemblage, not just game fish. The index incorporates professional judgment of fish assemblage health in 12 metrics and their scoring criteria, which are based on regional ideals. These regional standards are determined from historical data and data from minimally affected sites that characterize the region. The individual metrics differ in their range of sensitivity for detecting perturbations, and a degree of redundancy is built into the IBI because no single metric can reliably indicate integrity. The metrics and variations on them (Karr et al. 1986; Miller et al. 1988; Plafkin et al. 1989) are summarized below.

- Number of native fish species
- Number of darter and benthic species
- Number of sunfish and water column species
- Number of sucker and long-lived species
- Number of intolerant species
- Percent top carnivore individuals
- Percent insectivorous individuals
- Percent omnivorous individuals
- Percent tolerant individuals
- Total number of individuals
- Percent hybrid and exotic individuals
- Percent diseased individuals

Five of Karr's original metrics have frequently required modification for streams outside the Midwest

(Miller et al. 1988). It may be instructive to examine this process if IBI modification is needed in the southeastern United States. The numbers of benthic insectivorous species and sculpin species were used in New England and Oregon, respectively, where darters are depauperate or rare. Both substitutions contain species that use benthic habitats for reproduction and feeding, have small home ranges, and are sensitive to degradation. In the Southeast, madtoms might be added to this metric. The number of native minnow or water-column species replaced sunfish species as a measure of pool quality in Oregon and the Northeast, respectively. The species in both groups are sensitive to changes in pool or water-column habitats, and sunfishes are introduced or depauperate in both regions. Insectivorous cyprinids are a dominant trophic group in the Midwest, but where they are less dominant, Karr's original metric was changed to "all insectivores" or to "specialized invertebrate feeders." The proportion of individuals as green sunfish has been replaced frequently. This metric assesses the degree to which species that are tolerant to a variety of stressors dominate. Common carp, white sucker, creek chub (Miller et al. 1988), and the percent of individuals of the 12 most tolerant species (Ohio Environmental Protection Agency 1988) have all been used where green sunfish were inappropriate or insensitive. The percent of hybrids has also been problematic in several regions; introduced or exotic species were substituted in Oregon and Colorado. This modification may be appropriate in Florida also, as a measure of the breakdown of reproductive separation zoogeographically or simply as an indication of biological pollution.

There is growing interest in applying the IBI to water resource management. The IBI is presently being used by the Illinois Environmental Protection Agency, the Kentucky Cabinet for Natural Resources and Environmental Protection, the Ohio Environmental Protection Agency, and the Tennessee Valley Authority. Several other State agencies (Alabama Department of Environmental Management, Iowa Conservation Commission, Kansas Department of Wildlife and Parks, Nebraska Department of Environmental Control, New York Department of Environmental Conservation, North Carolina Division of Environmental Management, Oklahoma Department of Health, Vermont Department of Environmental Conservation, and Wisconsin Department of Natural Resources) and the National Park Service are testing the IBI for use as a monitoring tool. Ohio Environmental Protection Agency (1988) uses the IBI as a legal criterion in its water quality standards program, and the USEPA supports the

IBI for use in monitoring (U.S. Environmental Protection Agency 1988; Plafkin et al. 1989).

## Information Sources

- Hughes, R. M., and D. P. Larsen. 1988. Ecoregions: an approach to surface water protection. *J. Water Pollut. Control Fed.* 60:486-493.
- Hughes, R. M., D. P. Larsen, and J. M. Omernik. 1986. Regional reference sites: a method for assessing stream potentials. *Environ. Manage.* 10:629-635.
- Karr, J. R. 1981. Assessment of biotic integrity using fish communities. *Fisheries* 6:21-27.
- Karr, J. R., D. Fausch, P. L. Angermeier, P. R. Yant, and I. J. Schlosser. 1986. Assessing biological integrity in running waters: a method and its rationale. *Ill. Nat. Hist. Surv., Spec. Publ.* 5. 28 pp.
- Miller, D. L., P. M. Leonard, R. M. Hughes, J. R. Karr, P. B. Moyle, L. H. Schrader, B. A. Thompson, R. A. Daniels, K. D. Fausch, G. A. Fitzhugh, J. R. Gammon, D. B. Halliwell, P. L. Angermeier, and D. J. Orth. 1988. Regional applications of an index of biotic integrity for use in water resource management. *Fisheries* 13:12-20.
- Ohio Environmental Protection Agency. 1988. Biological criteria for the protection of aquatic life. Volumes I, II, III. Ohio Environmental Protection Agency, Columbus. 376 pp.
- Omernik, J. M. 1987. Ecoregions of the conterminous United States. *Ann. Assoc. Am. Geograph.* 77:118-125. (map scale 1:7,500,000)
- Plafkin, J. L., M. T. Barbour, D. D. Porter, S. K. Gross, and R. M. Hughes. 1989. Rapid bioassessment protocols for use in streams and rivers: benthic macroinvertebrates and fish. U.S. Environ. Prot. Agency EPA/444/4-89-001. 212 pp.
- U.S. Environmental Protection Agency. 1988. Report of the national workshop on instream biological monitoring and criteria. U.S. Environmental Protection Agency, Office of Water Regulations and Standards, Washington, D.C. 34 pp.

# Comparison of Instream Flow Methods for Western Virginia

by

Donald J. Orth

*Department of Fisheries and Wildlife Sciences  
Virginia Polytechnic Institute and State University  
Blacksburg, Virginia 24061*

and

Paul M. Leonard

*EA Engineering, Science, and Technology, Inc.  
Sparks, Maryland 21152*

Reliable methods are needed for recommending minimum instream flows to protect aquatic life because conflicts between instream and offstream uses of water will increase in the future, even in some water-rich regions. Water resource planning can help avoid these conflicts only if we know the acceptable magnitude and duration of low flows to maintain current instream values. For basinwide planning purposes, simple methods, which require little or no field investigations, are required. While some of the existing methods tend to be conservative, the degree to which they protect habitat for fish is seldom determined. We applied physical microhabitat models (PHABSIM) for nine target fish species in four streams in the upper James River basin, Virginia, to (1) identify optimum flows to protect the fish fauna, (2) investigate the relation between optimum flow and average discharge, and (3) compare our findings with recommendations based on simple discharge methods.

Microhabitat availability for riffle-dependent species was most limited at low flows, whereas microhabitat availability for pool-dependent species was most limited at high flows. At each study site there was a rapid increase in riffle habitat as discharge increased above zero. Optimum flow maximized habitat for the most critically habitat-limited species or life stages. The recommended optimum flows increased with increases in stream size, but the slope was not constant; as stream size increased, a lower proportion of average discharge provided optimum habitat. The recommended flows were related to the average discharge (AD) with a power function: optimum flow ( $m^3/s$ ) =  $0.583 AD^{0.746}$ .

Water resource managers can develop flow recommendations for other streams in the upper James River basin based on the average discharge—assuming water quality is not a limiting factor. Aquatic Base Flow recommendations (i.e., September median flow) provided varying—but a reasonable degree of—habitat protection. The Montana method (10% average discharge) recommendations correctly identified degraded or poor habitat conditions, and the 30% recommendations corresponded to near optimum habitat in small streams but greater than optimum flow at the large-stream site. Seven-day, 1-in-10-year low flows (7Q10) provided very limited amounts of physical habitat for riffle-dwelling fishes. The results of our study provide a basis for making preliminary flow recommendations in this region from readily available data. Studies will be needed, however, to test the generality of the findings in other basins and the assumptions of the methods employed.

## Information Sources

- Gore, J. A., and J. M. Nestler. 1988. Instream flow studies in perspective. *Regulated Rivers* 2:93–101.
- Leonard, P. M., and D. J. Orth. 1988. Use of habitat guilds of stream fishes to determine instream flow requirements. *N. Am. J. Fish. Manage.* 8:399–409.
- Leonard, P. M., D. J. Orth, and C. J. Goudreau. 1986. Development of a method for recommending instream flows for fishes in the upper James River, Virginia. *Va. Water Resour. Res. Cent. Bull.* 152. 122 pp.
- Lobb, M. D., III, and D. J. Orth. 1988. Microhabitat use by the bigmouth chub *Nocomis platyrhynchos* in the New River, West Virginia. *Am. Midl. Nat.* 120:32–40.

- Orth, D. J. 1981. Considerations in the development of curves for habitat suitability criteria. Pages 124–133 in N. B. Armantrout, ed. Symposium on acquisition and utilization of aquatic habitat inventory information. Western Division, American Fisheries Society, Bethesda, Md. 383 pp.
- Orth, D. J. 1987. Ecological considerations in the development and application of instream flow-habitat models. Regulated Rivers 1:171–181.
- Orth, D. J., and O. E. Maughan. 1982. Evaluation of the Incremental Methodology for recommending instream flows for fishes. Trans. Am. Fish. Soc. 111:413–445.
- Orth, D. J., and O. E. Maughan. 1986. In defense of the Instream Flow Incremental Methodology. Can. J. Fish. Aquat. Sci. 43:1092–1093.

## Minimum Flow is a Myth

by

Clair B. Stalnaker

*U.S. Fish and Wildlife Service  
National Ecology Research Center  
4512 McMurray Avenue  
Fort Collins, Colorado 80525*

During the warmwater stream symposium in 1980, when discussing low flow as a limiting factor in warmwater streams, I warned fisheries management personnel that the concept of a single minimum or base flow for fishery habitat maintenance that has evolved in the western region of the United States could very well become a real threat to low gradient, eastern, warmwater stream fisheries (Stalnaker 1981). The minimum flow concept rose from western water law as a mechanism to either reserve an amount of water from future appropriations or as a means of granting an instream water right for fishery purposes. This led to the myth that a consistent methodology could be used to establish a single minimum discharge value for any given stream. Experience has shown that as water becomes fully appropriated to upstream use or storage, the minimum flow, if not frequently violated in time, tends to become the average flow condition. Too often the minimum becomes the objective rather than the means to achieve some riverine fishery or recreation management goal. Such persistent low flows are not necessarily desirable from the water management perspective, being inflexible in the face of competitive uses or during unusual water supply conditions (e.g., drought), and certainly do not meet all the desired environmental needs. This difficulty with minimum flows arises in part because all the instream uses for which flows may be needed are not identified. Most often overlooked are necessary periodic high flows that move bedload, flush sediments, rejuvenate the floodplain, and generally maintain the structural characteristics of a stream channel, which should be maintained in dynamic equilibrium with its watershed (Stalnaker 1979).

A common misconception among water management personnel and consumers is that inclusion of all the identifiable instream uses of water in an instream flow requirement will dictate an additive treatment of their respective needs. This, it is fur-

ther assumed, will result in total allocation of the stream flow to instream uses. Contrary to this view, a considerable degree of compatibility exists among many instream uses and downstream delivery requirements for offstream or consumptive uses. However, in order to deal with these compatible uses, the instream flow advocate and the water resource manager must be aware of both the timing and the magnitude of all the demands being placed on the stream system. Such a common understanding, which should lead to the identification of instream flow requirements, will protect all complimentary uses as well as meet downstream delivery requirements.

It is evident from reviewing the literature and from the discussions during this workshop that many methods for evaluating instream flow needs have evolved since the 1960's. I prefer to categorize such methods as "standard setting" or "incremental." Standard setting methodologies, on one hand, refer to those measurements and interpretive techniques designed to generate a flow value (or values) intended to maintain the fishery or recreational use at some acceptable level (usually dictated by policy). Incremental methodologies, on the other hand, are organized and repeatable processes by which (1) a fishery habitat-stream flow relation and the hydrology of the stream are transformed into a baseline habitat time series, (2) proposed water management alternatives are simulated and compared with the baseline, and (3) project operating rules are negotiated.

Trihey and Stalnaker (1985) suggested that a hierarchical approach to hydro licensing and relicensing be followed that in essence takes advantage of both the standard setting and incremental approaches. A three-tiered hierarchy was suggested including reconnaissance, feasibility, and operational or design studies for evaluating hydro projects. It is important to recognize that such licensing is

generally a multiyear process. Adopting the suggested hierarchical approach can lead to greater understanding among the resource agencies, the applicant, and the general public, leading to negotiated conditions for the license. Specifically, the reconnaissance study identifies the stream segments of potential impact, the project location configuration, and possible operating scheme. With- and without-project hydrologic conditions are compared to determine whether the project seems to be "benign" and compatible with resource agency policies. In other words, there is little change in the flow pattern below the project. In the feasibility study, the use of a previously set standard can be quite advantageous. At this level of analysis, comparison is made between the projected stream flow conditions and the stream flow maintenance standard to identify major issues and periods of incompatibility. Standard setting methods (such as the New England Flow Method and the Arkansas and North Carolina methods) were discussed during this workshop; they and the optimum flow proposed for western Virginia are excellent examples by which one can screen for hydro projects that seem to be incompatible with agency policy and environmental protection goals. When it becomes obvious that project operations and the maintenance of stream flow standards are incompatible, impacts need to be quantified and mitigation measures agreed on. Then much more detailed operational level studies are appropriate. Only during this third study phase do the incremental methods become useful and, in fact, necessary.

The majority of States now recognize instream flows and have identified procedures for incorporating such uses in water planning (Reiser et al. 1989). Adoption of a standard setting approach by the State Water Resources and Fisheries Management agencies greatly facilitates identification of incompatible water development projects during feasibility studies. Stream flow assessment methods, such as the Instream Flow Incremental Methodology used by the U.S. Fish and Wildlife Service, have consequently evolved to become environmental assessment techniques and are used for evaluating the effects of proposed reservoir construction, water diversions, or hydroelectric operations on downstream fish habitats. Quite often such impact assessments become a matter of comparison among several possible, but not always measurable, water management schemes, leading to the necessity of simulation modeling for making these comparisons. Only the physical-chemical aspects of the habitat are evaluated, and comparisons are judged on the potential habitat limitations that may result from a proposed change in the way stream flows are controlled

and routed through stream segments. It is important to realize that minimum flows, optimal flows, and even stream flow standards are not impact assessment tools. When it comes to relicensing of hydroelectric projects, the questions really are focused on the effects that may result from a change in project operations. Minimum flow has no logical argument in such an institutional process and, in fact, as hydro projects go to increased peaking operations (involving daily and hourly rapid fluctuations in the tailwater releases), it is often the high flows that are of more concern from a biological standpoint than the low or minimum flows.

The challenge now before us is to progress beyond the minimum flow and even habitat impact assessment and to focus on scientific principles in understanding riverine systems. Management biologists must get involved with water management in riverine environments. By definition, management is a designed and directed change in a system. The improvement of basic understanding of ecology of our stream systems, coupled with the use of engineering tools and simulation modeling, provides an opportunity for fisheries to be enhanced downstream of the many hydroelectric projects coming up for relicensing in the 1990's. This will occur only if fishery managers and natural resource agencies do the designing and directing of the change in the operating systems, working hand-in-hand with the hydro project applicants and the Federal Energy Regulatory Commission.

## Information Sources

- Bovee, K. D. 1982. A guide to stream analysis using the Instream Flow Incremental Methodology. Instream Flow Information Paper 12. U.S. Fish Wildl. Serv., FWS/OBS 82/26. 248 pp.
- Bovee, K. D. 1985. Evaluation of the effects of hydropoeaking on aquatic macroinvertebrates using PHABSIM. Pages 236-241 in F. W. Olson, R. G. White, and R. H. Hamre, eds. Proceedings of the symposium on small hydropower and fisheries. American Fisheries Society, Bethesda, Md. 497 pp.
- Bovee, K. D. 1986. Development and evaluation of habitat suitability criteria for use in the Instream Flow Incremental Methodology. Instream Flow Information Paper 21. U.S. Fish Wildl. Serv., Biol. Rep. 86(7). 235 pp.
- Bovee, K. D., and J. R. Zuboy. 1988. Proceedings of a workshop on development and evaluation of habitat suitability criteria. U.S. Fish Wildl. Serv., Biol. Rep. 88(11). 407 pp.
- Connell, J. H., and W. P. Sousa. 1983. On the evidence needed to judge ecological stability or persistence. Am. Nat. 121(6):789-824.
- Estes, C. C. 1984. Evaluation of methods for recommending instream flows to support spawning by salmon. M.S. thesis, Washington State University, Pullman. 156 pp.

- Leonard, P. M., D. J. Orth, and C. J. Goudreau. 1985. Development of a method for recommending instream flows for fishes in the upper James River, Virginia. *Va. Water Resour. Res. Bull.* 152, 122 pp.
- Loar, J. M., and M. J. Sale. 1981. Analysis of environmental issues related to small-scale hydroelectric development. V. Instream flow needs for fishery resources. Oak Ridge National Laboratory ORNL/TM-7861, Oak Ridge, Tenn. 123 pp.
- McKinney, M. J., and J. G. Taylor. 1988. Western state instream flow programs: a comparative assessment. *Instream Flow Information Paper 18*. U.S. Fish. Wildl. Serv., Biol. Rep. 89(2). 78 pp.
- Orsborn, J. F., and C. H. Allman. 1976. Instream flow needs, Vols. I and II. American Fisheries Society, Bethesda, Md. 551 pp. and 657 pp.
- Orth, D. J. 1987. Ecological considerations in the development and application of instream flow-habitat models. *Regulated Rivers* 1:171-181.
- Prewitt, C. G. 1982. The effect of depth-velocity correlations on aquatic physical habitat usability estimates. Ph.D. dissertation, Colorado State University, Fort Collins. 83 pp.
- Raley, C., W. Hubert, and S. Anderson. 1988. Maintenance of flows downstream from water development projects in Colorado, Montana, and Wyoming. U.S. Fish. Wildl. Serv., Biol. Rep. 88(27). 73 pp.
- Reiser, D. W., M. P. Ramey, and T. R. Lambert. 1985. Review of flushing flow requirements in regulated streams. Pacific Gas and Electric Company, Department Engineering Research, San Ramon, Calif. 97 pp.
- Reiser, D. W., T. A. Wesche, and C. Estes. 1989. Status of instream flow legislation and practices in North America. *Fisheries* 14(2):22-29.
- Stalnaker, C. B. 1979. Myths concerning instream flows: a background to understanding instream uses. Pages 1-7 in P. S. Wassenberg, S. Olive, J. L. Demott, and C. B. Stalnaker, eds. Elements in negotiating stream flows associated with federal projects. *Instream Flow Information Paper 9*. U.S. Fish Wildl. Serv., FWS/OBS 79/03. 71 pp.
- Stalnaker, C. B. 1981. Low flow as a limiting factor in warmwater streams. Pages 192-199 in L. Krumholz, ed. Warmwater streams symposium. American Fisheries Society, Bethesda, Md. 422 pp.
- Stalnaker, C. B. 1982. Instream flow assessments come of age in the decade of the 1970's. Pages 119-141 in W. T. Mason, Jr., and S. Iker, eds. Research on fish and wildlife habitat. U.S. Environ. Prot. Agency EPA-600/8-82.022. 248 pp.
- Stalnaker, C. B., R. T. Milhous, and K. D. Bovee. 1989. Hydrology and hydraulics applied to fishery management in large rivers. Pages 13-30 in D. D. Dodge, ed. Proceedings of the international large river symposium. Can. Spec. Publ. Fish. Aquat. Sci. 106. 629 pp.
- Trihey, E. W., and C. B. Stalnaker. 1985. Evolution and application of instream flow methodologies to small hydropower development: an overview of the issues. Pages 176-183 in F. W. Olson, R. G. White, and R. H. Hamre, eds. Proceedings of the symposium on small hydropower and fisheries. American Fisheries Society, Bethesda, Md. 497 pp.
- Wesche, T. A., and P. A. Rechard. 1980. A summary of instream flow methods for fisheries and related research needs. *Eisenhower Consortium Bull.* 9. Water Resources Research Institute, University of Wyoming, Laramie. 122 pp.

## Considerations in Applying IFIM to Warmwater Streams<sup>1</sup>

by

John M. Nestler

*U.S. Army Engineers Waterways Experiment Station  
CEWES-ES-Q  
3909 Halls Ferry Road  
Vicksburg, Mississippi 39180*

Increased diversion of riverine flows in the southeastern United States for irrigation, municipal, and industrial water supply has resulted in an increased awareness of the importance of adequate instream flows for protecting and maintaining riverine fishes. One of the most commonly employed means of assessing the effects of river water diversion on warmwater streams is the U.S. Fish and Wildlife Service's Instream Flow Incremental Methodology (IFIM). Because the methodology was originally developed for coldwater streams, a number of factors should be considered in applying the IFIM to warmwater streams. Some primary considerations include differences in hydrology, channel geometry, and biology between coldwater and warmwater systems.

The hydrograph of coldwater streams used to develop the IFIM is heavily influenced by snowmelt or groundwater recharge, which tends to result in long-term gradually varying flow conditions. In contrast, warmwater streams are more influenced by rain runoff and can, therefore, exhibit complex hydrologic patterns that reflect daily, synoptic, or seasonal influences. Consequently, simple hydrologic variables, such as mean or median monthly flow (commonly used to describe coldwater streams), may not be as relevant to aquatic biota in warmwater systems. Indeed, variability in the hydrograph and concomitant variations in habitat may be important features of these ecosystems, required for organic matter transport, nutrient cycling, or successful completion of life stages.

Channel morphometry in the coldwater stream systems used to develop the IFIM was usually simple, composed primarily of pool-riffle complexes, and variety in habitat conditions (depth, velocity, and substrate) was more limited than in warmwater systems. In contrast, many warmwater streams have compound channels and may include backwaters, sloughs, braided channels, snags, and other channel features that make habitat conditions within these systems more difficult to describe using cross sections and relatively simple hydrologic methods.

The biology of many coldwater biota (primarily salmonids) is well known and documented. Many studies have been performed that describe salmonid habitat requirements in terms of substrate and channel flow conditions. In contrast, the biology of warmwater aquatic biota is less known and often undocumented in terms of variables used in instream flow studies. In fact, even the term *warmwater biota* may be misleading because it implies the existence of a distinct category of aquatic biota restricted to warmwater stream ecosystems. In reality, there may be any number of warmwater "biotic assemblages."

Coldwater and warmwater stream ecosystems differ substantially in species number. Small- and medium-sized coldwater streams are usually characterized by a relatively small number (1 to 10) of fish species and usually include only one or two sport fishes. It is usually not difficult to identify a target species or life stage to serve as the focal point of the study. In contrast, a warmwater stream of similar size may have 30 or 40 species of fishes, with several of them having commercial or sport-fishing importance. Consequently, it is often difficult to identify a suitable target species for the analysis. If the habitat requirements of a large number of species are evaluated, then assessment of impact is complicated because each life stage may have substantially different flow optima.

<sup>1</sup>The conclusions and interpretations presented in this paper were based on research conducted under the auspices of the Environmental and Water Quality Operations Studies and Environmental Impact Research Program of the U.S. Army Corps of Engineers by the Waterways Experiment Station. Permission was granted by the Chief of Engineers to publish this information.

Comparison of suitability curves for coldwater and warmwater fishes reveals that, generally, warmwater stream biota are characteristically habitat generalists, able to occupy a wider range of depth, velocity, and substrate conditions than coldwater biota. Consequently, an analysis using the Physical Habitat Simulation (PHABSIM) system of the IFIM generates habitat-discharge relations that are broad and flat (without sharp inflection points) for warmwater stream ecosystems. Such habitat-discharge relations are difficult to use in an assessment or management context. A similar analysis performed on a coldwater stream ecosystem generally produces more definitive results.

In summary, the basic assumption that stream flow is an important factor determining habitat quality for warmwater fishes is probably valid, although the precise manner in which river flow is related to habitat (or biomass) has not been described to the level of detail required for assessment of impact. Consequently, application of the IFIM to assess effects of river regulation or water withdrawal on warmwater fishes cannot be performed at a level of defensibility equivalent to that of coldwater stream applications until the unique hydrology and biology of these systems are described in greater detail and these findings incorporated into the IFIM.

## Information Sources

Bovee, K. D. 1982. A guide to stream habitat analysis

- using the Instream Flow Incremental Methodology. U.S. Fish Wildl. Serv., FWS/OBS-82/26. 248 pp.
- Bovee, K. D. 1986. Development and evaluation of habitat suitability criteria for use in the Instream Flow Incremental Methodology. U.S. Fish Wildl. Serv., FWS/OBS-86/7. 235 pp.
- Gore, J. A., and J. M. Nestler. 1988. Instream flow studies in perspective. *Regulated Rivers* 2:93-101.
- Mathur, D. L., W. H. Bason, E. J. Purdy, Jr., and C. A. Silver. 1984. A critique of the Instream Flow Incremental Methodology. *Can. J. Fish. Aquat. Sci.* 42:825-831.
- Milhous, R. T., D. L. Wegner, and T. J. Waddle. 1981. User's guide to the Physical Habitat Simulation System. U.S. Fish Wildl. Serv., FWS/OBS-81/43. 254 pp.
- Larimore, R. W., and D. D. Garrels. 1985. Assessing habitats used by warmwater stream fishes. *Fisheries* 10(2):10-16.
- Nestler, J. M., R. T. Milhous, and J. B. Layzer. 1989. Instream habitat modeling techniques. Pages 295-315 in G. E. Petts and J. A. Gore, eds. Alternatives in regulated river management. CRC Press, Inc., Boca Raton, Fla. 344 pp.
- Nestler, J. M., R. T. Milhous, J. Troxel, and J. A. Fritsch. 1986. Effects of flow alterations on trout, angling, and recreation in the Chattahoochee River, between Buford Dam and Peachtree Creek. Tech. Rep. E-86-10, U.S. Army Corps of Engineers, Waterways Experiment Station, Vicksburg, Miss. 105 pp.
- Orsborn, J. F., and C. H. Allman, editors. 1976. Instream flow needs, Vols. I and II. American Fisheries Society, Bethesda, Md. 551 pp. and 657 pp.
- Orth, D. J. 1987. Ecological considerations in the development and application of instream flow-habitat models. *Regulated Rivers* 1:171-181.
- Orth, D. J., and O. E. Maughan. 1982. Evaluation of the Incremental Methodology for recommending instream flows for fishes. *Trans. American Fisheries Society* 111:413-445.

## Determination of Instream Flow Needs at Hydroelectric Projects in the Northeast

by

Gordon W. Russell

*U.S. Fish and Wildlife Service  
400 Ralph Pill Marketplace  
22 Bridge Street  
Concord, New Hampshire 03301*

The Fish and Wildlife Service (Service) developed its New England Flow Method in 1981 in anticipation of reviewing numerous applications for new hydroelectric projects. At the time, there were about 10,000 existing dams in New England that were not being used for hydropower, but whose development was possible with the passage of the Public Utility Regulatory Policies Act of 1978. The New England Flow Method continues to be used by the Service, although its incorporation by the Federal Energy Regulatory Commission into hydroelectric project licenses and its effectiveness in protecting aquatic habitat have not been evaluated until now.

The New England Flow Method uses historical stream flow data to determine instream flow needs at a given hydroelectric project. The major biological assumption of the method is that instream flow equivalent to historical median discharge in August (base flow conditions) and during spawning and incubation periods will protect aquatic habitat to the degree that would be expected under a natural flow regime. Recommended flows are derived from historical flow data for the river on which the project is located. In certain circumstances, regional historical flow data are used, based on gaging information for unregulated rivers throughout New England. The regional median August flow is equivalent to  $0.07 \text{ m}^3/\text{s}/\text{km}^2$  of drainage area ( $0.5 \text{ ft}^3/\text{s}/\text{mi}^2$ ).

Using the New England Flow Method, the median August flow (Aquatic Base Flow) is recommended as the instream flow requirement that applies throughout the year unless higher flows are needed on a seasonal basis for migration, spawning, or egg incubation. The method also allows for project releases to equal inflow to the project area, when the latter are lower than the prescribed flows. Inflows lower than historical monthly median discharges would be expected during natural drought

conditions or when upstream projects are storing water for peaking or flow augmentation.

Since its inception in 1981, the New England Flow Method has been the basis for instream flow requirements at 157 hydroelectric projects in New England (77% of a total 205 projects authorized by the Federal Energy Regulatory Commission). At 48 projects (23%), the Commission used alternate means for determining instream flows, including requiring run-of-river operation, which obviates the need for minimum flows.

Lack of data on baseline and postproject conditions precludes direct observation of the level of habitat protection provided by flows derived from the New England Flow Method. However, an indirect measure can be found by examining the results of 14 instream flow studies. These studies were conducted by project proponents between 1981 and 1988 and used habitat-based techniques, including the Instream Flow Incremental Methodology. Flows supported by the results of the instream flow studies ranged from 63% below to 90% above those derived from the New England Flow Method (median 11.1% below the Aquatic Base Flow). In all but three cases, the instream flow studies gave results that were lower than corresponding values for Aquatic Base Flow. These results suggest that the New England Flow Method produces conservative results, favoring aquatic resource protection.

Review of numerous hydroelectric projects under prescribed deadlines requires an efficient approach. Faced with a high workload and limited resources, Service biologists often need to quickly develop recommendations for instream flows that will adequately protect fish and wildlife resources. The New England Flow Method can be applied with minimal information and no field studies. In addressing issues related to minimum flows at hydroelectric projects

in New England, the method appears to meet the Service's needs.

## Information Sources

EA Engineering, Science, and Technology, Inc. 1986. Instream Flow Methodologies. Electric Power Research Institute Research Project 2194-S, Palo Alto, Calif.

- 313 pp.  
Larsen, H. N. 1981. Interim regional policy for New England stream flow recommendations. U.S. Fish and Wildlife Service, Northeast Region, Newton Corner, Mass. 3 pp.  
Loar, J. M., and M. J. Sale. 1981. Analysis of environmental issues related to small-scale hydroelectric development. A. Instream flow needs for fishery resources. Oak Ridge National Laboratory, Environ. Sci. Div. Publ. 1829. 123 pp.

## Discharge versus Habitat in Steep-gradient Piedmont Streams

by

Jeffrey W. Foltz

*Department of Aquaculture, Fisheries, and Wildlife  
Clemson University  
Clemson, South Carolina 29634*

The focus of previous, ongoing, and future stream fish habitat research is the relation of the fish community to substrate composition, water depth, and water velocity. My basic premise has been that a diverse habitat will support a diverse fish community if water quality is suitable. After many years we have come to expect reductions in fish species diversity following catastrophic changes to a stream's habitat, such as channelization. Fishes of the Piedmont in the southeastern United States may be divided into three broad categories of habitat users: sculpins (*Cottus* spp.) and darters (*Etheostoma* spp. and *Percina* spp.), which prefer swift waters of riffles; minnows and suckers, which use open runs and midstream pools; and sunfishes (*Lepomis* spp.) and basses (*Micropterus* spp.), which seek cover near undercut banks and jams. These three broad categories of habitat are defined largely in terms of water velocity and depth. Substrate diversity is also important, not only because it provides a variety of spawning habitat for fishes, but also because it is the primary site of aquatic insect productivity.

The ratio of riffles to pools is thought to represent a fundamental characteristic of a stream ecosystem. A cross section or transect across a Piedmont stream or river frequently produces a series or mixture of small pools and riffles. Whereas a pool by itself might serve as a feeding station for a predatory fish, suitability of the pool depends on its proximity to a riffle in order to provide drifting insects. The original riffle-pool concept was theorized as a longitudinal phenomenon; however, a balance of riffles and pools at an acceptable level (e.g., at a ratio equal to that at median flow) might very well be necessary for maintenance of a diverse fish community.

Longitudinal increases in fish species diversity are well known. In my research, fish species diversity was predictable from substrate diversity, and substrate diversity increased downstream. Fish abundance (i.e., catch per unit effort) was positively

related to width, depth, and percent cobble (i.e., rubble) and negatively related to percent bedrock and percent silt and sand in the substrate. Currently, I am researching (1) the relation between discharge and diversity of velocity-depth combinations, (2) the relation between discharge and the riffle-pool ratio, (3) how relations 1 and 2 are related to conventional wetted perimeter-discharge curves, and (4) fish community diversity in relation to velocity-depth diversity and the riffle-pool ratio. Field methods involve placing numerous transects in a study section and measuring velocity, depth, and substrate at 1-m intervals along each transect. Subsequently, each interval is categorized into 1 of 13 categories, which include dry, 3 riffle categories, 5 pool categories, and 4 categories of runs. Velocity-depth diversity seems to gradually decline with reductions in discharge, and it collapses near the inflection point on wetted perimeter-discharge curves. This reduction is primarily due to reduction in the 4 categories of runs. The riffle-pool ratio remains generally flat with reductions in discharge, but collapses near the wetted perimeter-discharge inflection point.

### Information Sources

- Deacon, J. E., and W. G. Bradley. 1972. Ecological distribution of fishes of the Moapa River in Clark Co., Nevada. Trans. Am. Fish. Soc. 101:408-419.
- Fausch, K. D., C. L. Hawkes, and M. G. Parsons. 1988. Models that predict standing crop of stream fish from habitat variables: 1950-1985. U.S. For. Serv., Pac. Northwest Res. Stn. Gen. Tech. Rep. PNW-GTR-213. 52 pp.
- Foltz, J. W. 1982. Fish species diversity and abundance in relation to stream habitat characteristics. Proc. Annu. Conf. Southeast. Assoc. Fish Wildl. Agencies 36:305-311.
- Fraser, J. C. 1972. Regulated stream discharge for fish and other aquatic resources—an annotated bibliography. FAO Fish. Tech. Pap. 112. 103 pp.

- Gorman, O. T., and J. R. Karr. 1978. Habitat structure and stream fish communities. *Ecology* 59:507-515.
- Harrel, R. C., B. J. Davis, and T. C. Dorris. 1967. Stream order and species diversity of fishes in an intermittent Oklahoma stream. *Am. Midl. Nat.* 78:428-436.
- Hocutt, C. H., and J. R. Stauffer, Jr. 1980. Biological monitoring of fish. D.C. Heath Co., Lexington, Mass. 416 pp.
- Hynes, H. B. N. 1970. The ecology of running waters. University of Toronto Press, Ont. 555 pp.
- Karr, J. R. 1981. Assessment of biotic integrity using fish communities. *Fisheries* 6:21-27.
- Kuehne, R. A. 1962. A classification of streams, illustrated by fish distribution in an eastern Kentucky creek. *Ecology* 43:608-614.
- Mundie, J. H. 1974. Optimization of the salmonid nursery stream. *J. Fish. Res. Board Can.* 31:1827-1837.
- Sheldon, A. L. 1968. Species diversity and longitudinal succession in stream fishes. *Ecology* 49:193-198.
- Sprules, W. M. 1947. An ecological investigation of stream insects in Algonquin Park, Ontario. *Univ. Toronto Stud. Biol. Ser.* 56:1-81.
- Tramer, E. J., and P. M. Rogers. 1973. Diversity and longitudinal zonation in fish populations of two streams entering a metropolitan area. *Am. Midl. Nat.* 90:366-374.
- Whiteside, B. G., and R. M. McNatt. 1972. Fish species diversity in relation to stream order and physicochemical conditions in the Plum Creek drainage basin. *Am. Midl. Nat.* 88:90-101.

## Use of Multiple Methods for Instream Flow Recommendations—A State Agency Approach

by

Steven E. Reed and James S. Mead

*Division of Water Resources  
Aquatic Ecology Branch  
P.O. Box 27687  
Raleigh, North Carolina 27611*

North Carolina's waters range from the coldwater trout streams of the Appalachian Mountains to the meandering blackwater streams of the Coastal Plain. In between lie the coolwater streams and rivers of the Foothills and the Piedmont's broad, turbid rivers. The State's instream flow issues are also diverse. Many of these issues involve hydropower projects—either the existing facilities or the planning and design of new ones—and include peak power production, diversions of various lengths, and run-of-river projects that illegally peak. There are also concerns with existing reservoirs constructed for purposes such as municipal and industrial water supply, recreation, or irrigation. Many of these projects are old, and most do not have a required minimum flow. Direct withdrawals of water from streams also affect instream flow and aquatic habitat, including withdrawals for municipal and industrial water supply, irrigation, aquaculture, and thermal power plant cooling water.

The Division of Water Resources has been involved in instream flow studies since 1977. The division has conducted over 50 wetted perimeter studies and 24 Instream Flow Incremental Methodology (IFIM) studies, and it has made numerous flow recommendations using desktop methods. Every available opportunity is taken in project evaluation to address instream flow concerns. The State Dam Safety Law is applicable to all non-Federal dams higher than 15 feet that impound at least 10 acre-feet of water. This law is used to stipulate minimum flows from new dams unless they are located on intermittent streams. Hydropower projects must obtain a Certificate of Public Convenience and Necessity (CPCN) from the North Carolina Utilities Commission before construction. If the hydropower project will not be receiving a license or exemption from the Federal Energy Regulatory Commission, then the Division of Water Resources will attach its

instream flow recommendation to the Utilities Commission certificate. Instream flow recommendations may also be added as conditions to State 401 water quality certification and Federal 404 permits for projects that affect stream flow regimes.

Many factors are considered in selecting an instream flow assessment method. First, the value of the resource to be protected is considered. For example, a prime redbreast sunfish stream is handled differently than a severely degraded stream. The magnitude and duration of the project's impact are also evaluated by such factors as distance to the next major tributary, length of the diversion, or number of acres of potentially affected habitat. An attempt is made to determine whether the project will become controversial and what type of negotiations might take place. Other concerns include the manpower available for a study and how much time remains before a decision must be made.

The instream flow methods used can be separated into desktop and field methods. Desktop methods are used for projects with small impact or when a recommendation must be made quickly. A reconnaissance visit is usually made, and a recommendation is developed using one of several desktop methods. A previous recommendation may be ratioed by drainage area to a new location if there is an existing instream flow study site nearby and if the initial field visit indicates that the projects are similar. Review of hydrologic data may be used to develop a recommendation. Stream flow data from an appropriate U.S. Geological Survey gage are analyzed for mean annual flow and 7Q10 (lowest flow for 7 consecutive days, with a 10-year recurrence interval), as well as monthly means, medians, and lowest daily flows of record. These data are used to compare natural and proposed flow regimes. In areas where aquatic habitat is deemed minimal or nonexistent, the 7Q10 flow becomes the instream

flow recommendation. No flow recommendations have been less than the 7Q10 flow.

The New England Method, as adapted for use in North Carolina, uses the September median daily flow as the instream flow recommendation. The standard recommendation from the North Carolina Wildlife Resources Commission and the U.S. Fish and Wildlife Service is that a flow equal to the September median be provided, or a site-specific instream flow study must be conducted.

Field methods are used to develop instream flow recommendations when potential impacts are high, the stream contains a high quality fishery, or the project will involve peak power production. A site-specific study may be conducted by a developer to determine if an earlier desktop recommendation can be lowered or to provide answers to "what if" questions during negotiations. Field study methods used by the Division of Water Resources include wetted perimeter, regression model, incremental wetted surface area, and the IFIM. A State and Federal interagency team conducts a site visit and determines the most appropriate field methods to be used.

Wetted perimeter is one of the more frequently used field methods. Stream cross-sections, or transects, are selected to represent all habitat types contained in the stream reach of interest. The bottom profile of each transect is surveyed relative to a benchmark. Stream discharge and water surface elevations are measured at a minimum of three different flows so that a stage-discharge relation can be developed for each transect. Plots of transect wetted perimeter and stage are used to find the point of inflection (water surface elevation at which further reductions in stage result in large losses of wetted perimeter). The discharge corresponding to the water surface elevation of the point of inflection becomes the recommended flow for each transect. Through habitat mapping of the stream reach of interest, the percentage of the stream represented by each transect or the stream reach coefficient is determined. These stream reach coefficients and the sensitivity of each transect to stage reductions are used to weight each individual transect recommendation and develop the overall flow recommendation for the study site.

The wetted perimeter method is fairly inexpensive, and the time required for data collection can range from 2 days for a regulated stream to more than a year for remote sites on unregulated waters. Data analysis requires only 2 to 3 days. Because this method does not consider depth or velocity of water, any portion of the channel under water is considered habitat. Disadvantages of the method include subjective identification of point of inflection and the

flow recommendation representing only a single flow of the entire year. There is no way to evaluate the relation of flow to habitat at flows above or below the recommendation.

The regression model method is another field approach that was developed to provide a wetted perimeter recommendation. Numerous parameters from over 50 study sites across the State were analyzed for their correlation with the wetted perimeter recommendation at each site. To develop an instream flow recommendation, the model requires only mean annual flow, 7Q10, and average width at wetted area-discharge point of inflection. Validation analyses for new sites showed that the model predicts well for Piedmont streams. This method requires only one field visit to select and survey transects. No discharge measurements are needed, and the survey data can be collected during any season except during high water. The disadvantages are the same as for the wetted perimeter method.

The incremental wetted surface area method was developed as an improvement to the wetted perimeter method. Field data collection is the same as for the wetted perimeter method. For a given discharge, this method calculates the wetted surface area at each transect (wetted perimeter  $\times$  length of stream represented by the transect), and the procedure is repeated for all flows of interest. Plots of wetted surface area versus discharge are developed for each transect and the entire study site to indicate flow versus habitat relation. The point of inflection on a plot of wetted surface area for the overall study site is usually the recommended flow. Individual transect plots can be checked to determine the amount of wetted surface area at the recommended or other flows. The time needed to complete field data collection for this method is the same as for the wetted perimeter method; data analysis then requires 4 to 5 days. The main advantage of the incremental wetted surface area method is that it provides an indication of how wetted surface area changes over a wide range of flows. Its main disadvantage is that any immersed channel is considered habitat.

The IFIM is considered the state-of-the-art method for instream flow analyses and is widely accepted and used. It is used for projects that (1) are expected to have significant impacts, (2) may affect an outstanding fishery, or (3) are proposed as peaking hydropower producers. It is also used where complicated negotiations will be required to arrive at the recommended stream flow regime. Depth, velocity, substrate, and cover are all used in developing the flow versus habitat relation. The IFIM requires more field time and considerably more data analysis time than previously described field methods. Conse-

quently, staff resources from agencies will be high, even if much of the work is conducted by a private consulting firm.

The availability of several methodologies for evaluating and recommending instream flows provides a multitiered approach to solving instream flow problems. Each higher method offers a more refined recommendation but requires more investment of time and resources by the agency. The level at which the return no longer justifies the effort will differ for each instream flow situation. Agencies in

North Carolina try to select the approach that most effectively addresses a particular case. If the situation changes, there is nothing to preclude advancing to a more complex technique.

## Information Source

Reed, S. E., and J. S. Mead. 1988. Technical assistance throughout North Carolina's approach to recommending instream flows. Colorado State University, Instream Flow Chronicle 5:1-2.

## Stream Habitat Analysis and Instream Flow Assessment: A State-Federal Effort in Arkansas

by

Danny J. Ebert

*U.S. Forest Service  
P.O. Box 1270  
Hot Springs, Arkansas 71902*

Steve P. Filipek

*Arkansas Game and Fish Commission  
#2 Natural Resources Drive  
Little Rock, Arkansas 71202*

and

Kelly M. Russell

*U.S. Forest Service  
1765 Highland Avenue  
Montgomery, Alabama 36107*

Recent legislation passed by the Arkansas Legislature requires the determination of instream flow requirements for beneficial uses in the State's major rivers. In response to this mandate, the Arkansas Game and Fish Commission and State Department of Pollution Control and Ecology developed the Arkansas Method of instream flow determination for fish and wildlife needs. The Arkansas Method is a modification of the Tennant Method (Tennant 1975, 1976) for instream flow assessment; it uses flow standards for three seasonal periods. Sixty percent of the mean monthly flow (MMF) for November through March is recommended for flushing sediments and shaping channels. Seventy percent of the MMF for April, May, and June is recommended to protect spawning. Fifty percent of the MMF (or median, in some cases) for July to October is recommended to maintain water quality and provide habitat conditions conducive to fish growth and production. The flow standards were developed from information on historic streamflows in Arkansas, field experience and data on stream fishes, and knowledge of natural seasonal processes.

Because many of the State's highest quality streams are found on National forests, a cooperative

State-Federal study was developed to assess stream flows in relation to fish population abundance, habitat, and water chemistry. Field studies were conducted from 1985 to 1988 in third- through fifth-order streams in the Ozark-St. Francis and Ouachita National Forests. Stream reaches were delineated by channel morphology and substrate, and entire pools or riffles were sampled at least three times each year, corresponding to Arkansas seasonal flows.

The structure of the fish community in each habitat type was summarized using feeding guilds, percent composition per species, and biomass per family group. Stream habitat was surveyed at each site using 24 habitat measurements, and water quality and instream flow measurements were made. The number of species generally increased from headwaters to midreaches and was in most cases associated with addition of new species rather than replacement. The majority of species added with increasing stream order were pool species or slow-water, large-channel species. Habitat diversity increased with downstream progression, canopy closure decreased as channels became wider, substrate particle size became more heterogenous, and

mean riffle-pool depth increased. Pool-riffle ratio in most Ozark and Ouachita streams remained constant (1:1) with progression to midreaches.

The studies were initiated to determine if flow recommendations computed using the Arkansas Method were appropriate for smaller-order streams of the Ozark-St. Francis and Ouachita National Forests. The Arkansas Method was found to be adequate to protect the stream fisheries in National forest watersheds. However, for streams that regularly have only subsurface flow during dry periods (i.e., intermittent as measured by streamflow gages), the Arkansas Method may need to be refined.

## Information Sources

- Baker, J. A., and S. T. Ross. 1981. Spatial and temporal resource utilization by southeastern cyprinids. *Copeia* 178:189.
- Barila, T. Y., R. D. Williams, and J. R. Stauffer, Jr. 1981. The influence of stream order and selected stream bed parameters on fish diversity in Raystown Branch, Susquehanna River drainage, Pennsylvania. *J. Appl. Ecol.* 18:125-131.
- Brussock, P. P., A. V. Brown, and J. C. Dixon. 1985. Channel form and stream ecosystem models. *Water Resour. Bull.* 21:859-866.
- Deacon, J. E., and W. G. Bradley. 1972. Ecological distribution of fishes of the Moapa River in Clark County, Nevada. *Trans. Am. Fish. Soc.* 101:408-419.
- Ebert, D. J., S. P. Filipek, and R. D. Patton. 1987. Guide to the monitoring and analysis of Arkansas streams. U.S. Forest Service, Russellville, Ark. 197 pp.
- Evans, J. W., and R. L. Noble. 1979. The longitudinal distribution of fishes in east Texas streams. *Am. Midl. Nat.* 101:333-343.
- Felley, J. D., and L. G. Hill. 1983. Multivariate assessment of environmental preferences of cyprinid fishes of the Illinois River, Oklahoma. *Am. Midl. Nat.* 109:209-221.
- Filipek, S. P., W. E. Keith, and J. Giese. 1987. The status of the instream flow issue in Arkansas. *Proc. Ark. Acad. Sci.* 41:43-48.
- Foltz, J. W. 1982. Fish species diversity and abundance in relation to stream habitat characteristics. *Proc. Annu. Conf. Southeast. Assoc. Fish Wildl. Agencies* 36:305-312.
- Garman, G. C., T. L. Thorn, and L. A. Nielson. 1982. Longitudinal variation of the fish community of Brumley Creek, Virginia, and implications for sampling. *Proc. Annu. Conf. Southeast. Assoc. Fish Wildl. Agencies* 36:386-394.
- Gorman, O. T., and J. R. Karr. 1978. Habitat structure and stream fish communities. *Ecology* 59:507-515.
- Guillory, V. 1982. Longitudinal gradients of fishes in Thompson Creek, Louisiana. *Southwest. Nat.* 27:107-115.
- Hankin, D. G. 1986. Sampling designs for estimating the total number of fish in small streams. *U.S. For. Serv. PMN-360.* 38 pp.
- Matthews, W. J. 1982. Small fish community structure in Ozark streams: structures assembly patterns or random abundance of species? *Am. Midl. Nat.* 107:42-54.
- Matthews, W. J., and L. G. Hill. 1979. Age specific differences in the distribution of red shiners (*Notropis lutrensis*) over physicochemical ranges. *Am. Midl. Nat.* 101:366-372.
- Matthews, W. J., and L. G. Hill. 1979. Influence of physicochemical factors on habitat selection by red shiners *Notropis lutrensis* (Pisces, Cyprinidae). *Copeia* 1979:70-81.
- Matthews, W. J., and L. G. Hill. 1980. Habitat partitioning in the fish community of a southwestern river. *Southwest. Nat.* 25:57-66.
- Matthews, W. J., and J. D. Maness. 1979. Critical thermal maxima, oxygen tolerances, and success of cyprinid fishes in a southwestern river. *Am. Midl. Nat.* 102:374-377.
- Matthews, W. J., and H. W. Robison. 1988. The distribution of the fishes of Arkansas: a multivariate analysis. *Copeia* 1988:358-374.
- Orth, D. J., and O. E. Maughan. 1984. Community structure and seasonal changes in standing stocks of fish in a warmwater stream. *Am. Midl. Nat.* 112:369-378.
- Platts, W. S., C. Armour, G. D. Booth, M. Bryant, J. L. Bufford, G. W. Linkaemper, G. W. Minshall, S. B. Monsen, R. L. Nelson, J. R. Sedell, and J. S. Tuhy. 1987. Methods for evaluating riparian habitats with applications to management. *U.S. For. Serv., Intermountain Res. Stn. Gen. Tech. Rep. INT-221.* 177 pp.
- Platts, W. S., W. F. Megahan, and G. W. Minshall. 1983. Methods for evaluating stream riparian and biotic conditions. *U.S. For. Serv., Intermountain Res. Stn. Gen. Tech. Rep. INT-38.* 70 pp.
- Rose, D. R., and A. A. Echelle. 1981. Factor analysis of association of fishes in Little River, central Texas, with a interdrainage comparison. *Am. Midl. Nat.* 106:379-391.
- Sheldon, A. L. 1968. Species diversity and longitudinal succession in stream fishes. *Ecology* 49:193-198.
- Smith, C. L., and C. R. Powell. 1971. The summer fish community of Brier Creek, Marshall County, Oklahoma. *Am. Mus. Novit.* 2458:1-30.
- Tennant, D. L. 1975. Instream flow regimens for fish, wildlife, recreation and related environmental resources. *U.S. Fish and Wildlife Service, Billings, Mont.* 30 pp.
- Tennant, D. L. 1976. Instream flow regimens for fish, wildlife, recreation and related environmental resources. *Fisheries* 1(4):6-10.
- VanDeventer, J. S., and W. S. Platts. 1985. A computer software system for entering, managing, and analyzing fish capture data from streams. *U.S. For. Serv., Intermountain Res. Stan. Res. Note INT-352.* 12 pp.
- Whiteside, B. G., and R. M. McNatt. 1972. Fish species diversity in relation to stream order and physicochemical conditions in the Plum Creek drainage basin. *Am. Midl. Nat.* 88:90-101.

Bain, Mark A., editor. 1990. **Ecology and Assessment of Warmwater Streams: Workshop Synopsis.** U.S. Fish Wildl. Serv., Biol. Rep. 90(5). 44 pp.

Natural resource agencies are frequently confronted by applications for actions that could impact aquatic resources of warmwater streams and rivers. To assess impacts and develop mitigation measures, agency biologists need state-of-the-art knowledge on warmwater stream ecology and methods for predicting changes. A 1988 workshop reviewed selected topics on stream ecology and impact assessment. Summaries of 18 presentations by basic and applied researchers introduce recent information and methods. Research on the ecology of warmwater streams emphasized four common themes: understanding causes of natural variation in biological communities, importance of the magnitude and timing of streamflow changes, relations between early life stages and fish population size, and patterns of stream habitat use at the community level. Applied research biologists assessing impacts to streams varied in their approach. One group advocates the use of readily obtainable data and emphasizes assessment methods with narrow, specific objectives. Another group advocates techniques based on communities and streamflow regimes and they believe that stream protection will not be achieved by focusing on one or a few species and a single standard for stream protection. Deficiencies in knowledge of stream ecology and inadequacies in available impact assessment methods indicate that much more basic and applied research will be needed before warmwater streams can be confidently managed.

**Key words:** Stream ecology, impact assessment, habitat requirements, instream flow, macroinvertebrates, fish communities.

Bain, Mark A., editor. 1990. **Ecology and Assessment of Warmwater Streams: Workshop Synopsis.** U.S. Fish Wildl. Serv., Biol. Rep. 90(5). 44 pp.

Natural resource agencies are frequently confronted by applications for actions that could impact aquatic resources of warmwater streams and rivers. To assess impacts and develop mitigation measures, agency biologists need state-of-the-art knowledge on warmwater stream ecology and methods for predicting changes. A 1988 workshop reviewed selected topics on stream ecology and impact assessment. Summaries of 18 presentations by basic and applied researchers introduce recent information and methods. Research on the ecology of warmwater streams emphasized four common themes: understanding causes of natural variation in biological communities, importance of the magnitude and timing of streamflow changes, relations between early life stages and fish population size, and patterns of stream habitat use at the community level. Applied research biologists assessing impacts to streams varied in their approach. One group advocates the use of readily obtainable data and emphasizes assessment methods with narrow, specific objectives. Another group advocates techniques based on communities and streamflow regimes and they believe that stream protection will not be achieved by focusing on one or a few species and a single standard for stream protection. Deficiencies in knowledge of stream ecology and inadequacies in available impact assessment methods indicate that much more basic and applied research will be needed before warmwater streams can be confidently managed.

**Key words:** Stream ecology, impact assessment, habitat requirements, instream flow, macroinvertebrates, fish communities.

Bain, Mark A., editor. 1990. **Ecology and Assessment of Warmwater Streams: Workshop Synopsis.** U.S. Fish Wildl. Serv., Biol. Rep. 90(5). 44 pp.

Natural resource agencies are frequently confronted by applications for actions that could impact aquatic resources of warmwater streams and rivers. To assess impacts and develop mitigation measures, agency biologists need state-of-the-art knowledge on warmwater stream ecology and methods for predicting changes. A 1988 workshop reviewed selected topics on stream ecology and impact assessment. Summaries of 18 presentations by basic and applied researchers introduce recent information and methods. Research on the ecology of warmwater streams emphasized four common themes: understanding causes of natural variation in biological communities, importance of the magnitude and timing of streamflow changes, relations between early life stages and fish population size, and patterns of stream habitat use at the community level. Applied research biologists assessing impacts to streams varied in their approach. One group advocates the use of readily obtainable data and emphasizes assessment methods with narrow, specific objectives. Another group advocates techniques based on communities and streamflow regimes and they believe that stream protection will not be achieved by focusing on one or a few species and a single standard for stream protection. Deficiencies in knowledge of stream ecology and inadequacies in available impact assessment methods indicate that much more basic and applied research will be needed before warmwater streams can be confidently managed.

**Key words:** Stream ecology, impact assessment, habitat requirements, instream flow, macroinvertebrates, fish communities.

Bain, Mark A., editor. 1990. **Ecology and Assessment of Warmwater Streams: Workshop Synopsis.** U.S. Fish Wildl. Serv., Biol. Rep. 90(5). 44 pp.

Natural resource agencies are frequently confronted by applications for actions that could impact aquatic resources of warmwater streams and rivers. To assess impacts and develop mitigation measures, agency biologists need state-of-the-art knowledge on warmwater stream ecology and methods for predicting changes. A 1988 workshop reviewed selected topics on stream ecology and impact assessment. Summaries of 18 presentations by basic and applied researchers introduce recent information and methods. Research on the ecology of warmwater streams emphasized four common themes: understanding causes of natural variation in biological communities, importance of the magnitude and timing of streamflow changes, relations between early life stages and fish population size, and patterns of stream habitat use at the community level. Applied research biologists assessing impacts to streams varied in their approach. One group advocates the use of readily obtainable data and emphasizes assessment methods with narrow, specific objectives. Another group advocates techniques based on communities and streamflow regimes and they believe that stream protection will not be achieved by focusing on one or a few species and a single standard for stream protection. Deficiencies in knowledge of stream ecology and inadequacies in available impact assessment methods indicate that much more basic and applied research will be needed before warmwater streams can be confidently managed.

**Key words:** Stream ecology, impact assessment, habitat requirements, instream flow, macroinvertebrates, fish communities.

---

**NOTE:** The opinions and recommendations expressed in this report are those of the authors and do not necessarily reflect the views of the U.S. Fish and Wildlife Service, nor does the mention of trade names constitute endorsement or recommendation for use by the Federal Government.

# TAKE PRIDE *in America*



U.S. DEPARTMENT OF THE INTERIOR  
FISH AND WILDLIFE SERVICE



As the Nation's principal conservation agency, the Department of the Interior has responsibility for most of our nationally owned public lands and natural resources. This includes fostering the wisest use of our land and water resources, protecting our fish and wildlife, preserving the environmental and cultural values of our national parks and historical places, and providing for the enjoyment of life through outdoor recreation. The Department assesses our energy and mineral resources and works to assure that their development is in the best interests of all our people. The Department also has a major responsibility for American Indian reservation communities and for people who live in island territories under U.S. administration.